Antralization of the Gastric Mucosa of the Incisura Angularis and its Gastrin Expression

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Abstract: The frequency of antrum-type mucosa and gastrin expression in gastric biopsies from the incisura angularis was assessed in 60 consecutive patients having gastrointestinal symptoms. Following the recommendations from the updated Sydney System for the classification and grading of gastritis, two biopsies were taken from the antrum, one from the incisura and two from the corpus. Sections were stained with H&E, Giemsa and for gastrin. Gastrin-positive cells were semi-quantified as: 0 (none), ≤ 9, 10-49 cells and ≥50 gastrin-labelled cells/40x field. Antrum-type mucosa at the incisura (called antralization) occurred in 30% of the biopsies without inflammation, but in 69% of those with H. pylori-induced gastritis, and in 64% of those with autoimmune gastritis. At the incisura, gastrin-labelled cells (≥10) were found in 62% (18/29) of biopsies showing antralization, but in only 20% (3/15) of those having transitional-type mucosa (p<0.05) and in none of the 16 biopsies having fundic-type mucosa. The similarity in gastrin expression between the mucosa of the gastric antrum and the antral-type mucosa at the incisura substantiates the notion that antralization is a metaplastic transformation. The significantly higher frequency of antral-type mucosa at the incisura in patients with gastritis than in those without gastritis strongly suggests that chronic inflammation per se triggers antralization of the incisura, irrespective of the presence or absence of H. pylori infection.

Key Words: Antralization, incisura, gastrin, chronic gastritis

Introduction

The stomach is anatomically subdivided into four segments: the cardia, the fundus, the corpus or body and the antrum. The frontal profile of the stomach is delineated by the lesser and the greater curvatures [1]. The lesser curvature extends from the cardia to the pyloric sphincter and displays an abrupt change in spatial direction from vertical to horizontal. The vertical (proximal) section limits the medial aspect of the fundus-corpus and the horizontal (distal), the medial aspect of the antrum pylori. The angle in the lesser curvature moulded by the vertical and the horizontal sections is called incisura angularis [1].

The fundic (fundus-corpus) mucosa is built of parietal cells/chief cells while the mucosa of the antrum is built of mucin-producing cells and specialized endocrine cells. The mucosa of the incisura angularis is comprised of an admixture of corpus and antral mucosa [2], hence intermediate mucosa.

About 50% of the entire endocrine cell population in the antrum are gastrin-producing (G cells). G cells are most common in the neck region just below the pits [2]. Gastrin antibodies detect cytoplasmic granules located between the nucleus and the basement membrane. Gastrin is produced in response to stimuli such as stomach distension, vagal stimulation (mediated by bombesin), presence of luminal amino acids and hypercalcemia. The release of gastrin is inhibited by increased intragastric acidity. Similar endocrine cells are not present in the fundic mucosa [2-4]. Antral gastrin stimulates parietal cells to produce
hydrochloric acid (HCl). This is done indirectly by binding CCK2/gastrin receptors on endocrine-like cells (ELC) through the release of histamine. Acting in a paracrine fashion, parietal cells are stimulated to secrete H+ ions. Gastrin also causes chief cells to secrete pepsinogen, which is converted into pepsin under the influence of a high HCL production and of pancreatic enzymes from acinar cells [2-4].

Endoscopical gastric biopsies are usually taken in symptomatic patients to assess the histological state of the gastric mucosa. This method permits to diagnose gastritis (grade of inflammation), Helicobacter pylori infection and gastritis sequels, such as mucosal atrophy, pseudopyloric metaplasia and intestinal metaplasia [5-18].

Four biopsies are usually taken, two from the antrum and two from the corpus. The updated Sydney System for the classification and grading of gastritis [19] recommends to include, in addition to the four aforementioned sites, a fifth biopsy from the incisura angularis.

More recently, Xia et al [20-23] studied mucosal changes occurring at the incisura angularis. They classified the mucosa of the incisura angularis into i) antral-type, with mucous-secreting pyloric glands (called antralization of the mucosa of the incisura); ii) transitional-type with mixed antrum-body-glands (called junctional mucosa) and corpus/body-type with acid-secreting or oxyntic glands.

Gastrin is a marker of the pyloric mucosa [2]. No studies have been carried out to explore the presence of this hormone in the various mucosal subtypes occurring at the incisura angularis [20-23]. In this work, we investigated the occurrence of gastrin-producing cells in the antrum, the incisura and the corpus, with particular attention to the presence of gastrin-expressing cells in the various mucosal subtypes at the incisura angularis.

Material and Methods

Specimen Collection

Gastric biopsies were obtained in 60 patients having gastrointestinal symptoms. Patients receiving H2 receptor blockers, on PPI therapy or on nonsteroid anti-inflammatory drug (NSAID) at entry were excluded. The Regional Ethical Committee approved the study.

Gastric biopsies were taken from 5 different sites, following the recommendations from the updated Sydney System for the classification and grading of gastritis [19]: two from the antrum, one from the incisura angularis and two from the corpus.

Special Stains

Biopsies were fixed in 4% buffered formalin and processed for paraffin embedment. Sections were routinely stained with hematoxylin-eosin (H&E) and Giemsa. Immunohistochemical stain with gastrin (DAKO, Glistrup, Denmark) antibody (dilution at 1:4000) was performed using the standard protocol with DAB as a detector and appropriate positive control.

A high-power field (40x) containing a priori most gastrin-labelled cells was chosen to assess the number of gastrin-positive mucosa cells. The frequency of gastrin-positive cells/40x field was semi-quantified as follows: no labelled cells: 0; ≤ 9 labelled cells; 10-49 labelled cells and ≥50 labelled cells.

After grading the immunoreactions, the histological make-up of the incisura angularis was classified in H&E sections, blind to the results of the immunohistochemical reactions into the 3 subtypes proposed by Xia et al [20, 21, 23].

Statistical Analysis

The non-parametric test of Wilcoxon was performed using StatView Version 4.5 software (Abacus Concepts, Berkley, CA, USA). Statistical significance was defined as p<0.05.

Results

Among the 60 patients, 33 were females and 27 were males. The mean age was 58.6 years, ranging from 18 to 89 years.

Histological examination revealed that 30 patients had normal mucosa, 16 had H. pylori-induced gastritis and 14 had autoimmune gastritis (Table 1). Autoimmune gastritis is an inflammatory condition of the stomach that is associated with autoantibodies to parietal cells and intrinsic factor and can lead in a small
Rubio C et al/Gastrin Expression in Gastric Biopsies

Table 1  Antralization of the mucosa at the incisura angularis

<table>
<thead>
<tr>
<th>Mucosa at the incisura angularis</th>
<th>Antral-type</th>
<th>Transitional-type</th>
<th>Corpus-type</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>9 (30%)</td>
<td>11 (36.7%)</td>
<td>10 (33.3%)</td>
<td>30 (100%)</td>
</tr>
<tr>
<td><em>H. Pylori</em> Gastritis</td>
<td>11 (68.8%)</td>
<td>2 (12.5%)</td>
<td>3 (18.7%)</td>
<td>16 (100%)</td>
</tr>
<tr>
<td>Autoimmune gastritis</td>
<td>9 (64.3%)</td>
<td>2 (14.3%)</td>
<td>3 (21.4%)</td>
<td>14 (100%)</td>
</tr>
<tr>
<td>Total</td>
<td>29 (48.3%)</td>
<td>15 (25.0%)</td>
<td>16 (26.7%)</td>
<td>60 (100%)</td>
</tr>
</tbody>
</table>

percentage of patients to destruction of the oxyntic mucosa, pernicious anemia, and the development of carcinoid tumors that are typically indolent. Autoimmune gastritis is also referred to as Type A gastritis and is recognized as a corpus-restricted atrophic gastritis in the updated Sydney classification system [19].

The mean age in the 30 patients having histologically normal gastric mucosa was 56.6 years (range: 18-80 years), 60.8 years (range 27-89) in the 16 patients with *H. pylori* gastritis, and 64.1 years (range 26-89 years) in the 14 patients with autoimmune gastritis. The difference was not significant (p< 0.6).

The mean age of patients having antral-type mucosa at the incisura was 63.2 years (range 27-89 years), 52.2 years (range 26-76 years) in patients having transitional-type mucosa and 60.0 years (range 35-89 years) for those having fundic-type mucosa. Although the mean age for patients having antral mucosa at the incisura was higher than those having transitional-type mucosa, the difference was not significant (p<0.6).

The histological finding of mucosa at the incisura is summarized in Table 1. Among the 60 biopsies from the incisura, 48% had antrum-type mucosa only, 25% had intermediate-type mucosa and the remaining 27% had fundic-type mucosa only.

Antralization of the incisura occurred in 30% of the 29 patients having normal gastric mucosa, in 69% of the 16 patients with *H. pylori*-induced gastritis, and in 64% of the 14 patients with autoimmune gastritis (Table 1). The difference between antralization of the incisura angularis in patients without inflammation and those with inflammation (*H. pylori*-induced gastritis and autoimmune gastritis) was significant (p<0.05).

The frequency of gastrin-labelled cells in the mucosa of the antrum and body was summarized in Table 2. Among the 60 antral biopsies, 45% had ≥50 gastrin-labelled cells (Figure 1), 50% had 10 to 49 gastrin-labelled cells, and the remaining 5% had ≤9 gastrin-labelled cells. In contrast, only 6 or 10% of the 60 biopsies from the fundic mucosa contained ≤9 gastrin-labelled cells. The 6 biopsies were all from patients with autoimmune gastritis. The remaining 54 biopsies from the fundic mucosa showed no gastrin-labelled cells.

### Table 2  Gastrin expression in the mucosa of the antrum and body

<table>
<thead>
<tr>
<th>Histological phenotype</th>
<th>No labelled cells</th>
<th>≤ 9 labelled cells</th>
<th>10-49 labelled cells</th>
<th>≥50 labelled cells</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Antrum</td>
<td>0</td>
<td>3 (5.0%)</td>
<td>30 (50%)</td>
<td>27 (45%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Body</td>
<td>54 (96.0%)</td>
<td>6 (10.0%)</td>
<td>0</td>
<td>0</td>
<td>60 (100%)</td>
</tr>
</tbody>
</table>
Figure 1 Diagram of the stomach anatomy (middle panel) with positive and negative expression of gastrin in the antrum and corpus, respectively (right panel). Representative images showing strong expression of gastrin in the antrum-type mucosa (A), discontinuous expression of gastrin in the transitional-type mucosa (B) and absent expression of gastrin in the fundic-type mucosa at the incisura (C). (gastrin immunostains, all at 4X magnifications).

Table 3 Gastrin expression in the mucosa of the incisura angularis

<table>
<thead>
<tr>
<th>Gastrin</th>
<th>Antral-type</th>
<th>Transitional-type</th>
<th>Fundic-type</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>No labelled cells</td>
<td>2</td>
<td>1</td>
<td>16</td>
<td>19 (31.7%)</td>
</tr>
<tr>
<td>≤ 9 labelled cells</td>
<td>9</td>
<td>11</td>
<td>0</td>
<td>20 (33.3%)</td>
</tr>
<tr>
<td>10-49 labelled cells</td>
<td>8</td>
<td>2</td>
<td>0</td>
<td>10 (16.7%)</td>
</tr>
<tr>
<td>≥50 labelled cells</td>
<td>10</td>
<td>1</td>
<td>0</td>
<td>11 (18.3%)</td>
</tr>
<tr>
<td>Total</td>
<td>29 (48.3%)</td>
<td>15 (25.0%)</td>
<td>16 (26.7%)</td>
<td>60 (100%)</td>
</tr>
</tbody>
</table>

5% (n=3) of the 60 biopsies from the antrum and 10% (n=6) from the fundic mucosa had ≤9 gastrin-labelled cells, whereas 38% (n=11) of the 29 biopsies from the incisura with antrum-type mucosa had ≤9 gastrin-labelled cells. The difference was significant (p<0.05).

The frequency of gastrin-labelled cells in the mucosa of the incisura is summarized in Table 3. Among the 29 biopsies from the incisura with antrum-type mucosa, 34% (n=10) had ≥50 gastrin-labelled cells. In contrast, only 7% (n=1) of the 15 biopsies from the incisura with transitional-type mucosa had ≥50 gastrin-labelled cells and in none (=0) of the 16 biopsies with fundic-type mucosa at the incisura (p<0.05) (Figure 1). Similarly, 62% (n=18) of the 29 biopsies from the incisura having antrum-type mucosa had ≥10 gastrin-labelled cells, while only 20% (n=3) of the 15 biopsies from the incisura having transitional-type mucosa had ≥10 gastrin-labelled cells (p<0.05). None of the 16 biopsies from the incisura having fundic-type mucosa had gastrin-labelled cells.
Discussion

Several authors have investigated the relationship between the mucosa of the incisura angularis and *H. pylori* infection. Dursun et al. [25] reported that the highest sensitivity in detecting *H. pylori* infection occurred in biopsies taken from the antrum and the incisura. On the other hand, Ola et al. [26] found no benefit in taking additional biopsies from the incisura angularis in assessing *H. pylori* infection. Sheng-Lian et al. [15] reported that the fundic mucosa at the incisura from patients with bile reflux had more severe active inflammation, chronic inflammation, intestinal metaplasia, atrophy or *H. pylori* infection than in control patients without reflux. Stolte et al. [24] found that 58% of 328 gastric biopsies from the incisura contained antral mucosa, 24% intermediate mucosa and 18% corpus mucosa. According to these authors, biopsies showing antral mucosa were exclusively associated with more severe *H. pylori* gastritis in both the incisura and the antrum, than in those having corpus mucosa only. Xia et al. [20, 21, 23] investigated antralization of the incisura angularis and of the body mucosa (known as pseudopyloric metaplasia of the body). These authors [20, 21] claimed that under the influence of *H. pylori*, the hyperplastic mucous neck cells moved both upwards and particularly downwards in the oxyntic tubules, replacing the specialized parietal and chief cells in the body and the incisura, creating a mucous cell lineage (ulcer-associated cell lineage, UACL).

Similar other studies [21-23], we found here that the histological make-up of the mucosa of the incisura varied. 48% had antral-type mucosa exclusively, 25% had intermediate mucosa and the remaining 27% had corpus-type mucosa exclusively.

Antralization at the incisura concurred with a significantly higher expression of gastrin-producing cells, than intermediate and fundic-type mucosas. The similarity in gastrin-expression between the gastric mucosa of the antrum and the antral-type mucosa at the incisura strongly suggests that antralization of the incisura is conveyed by metaplastic transformation.

Metaplasia is usually defined as the reversible replacement of one differentiated cell type by another mature differentiated cell type. Whether the metaplastic antralization of the mucosa of the incisura is reversible remains to be elucidated.

The frequency of antralization of the incisura was similarly high in patients with *H. pylori* gastritis and with autoimmune gastritis. These results strongly suggest that chronic inflammation per se plays a causative role in the antralization of the gastric mucosa at the incisura angularis, irrespective of the presence or absence of *H. pylori* infection.

Acknowledgement

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