Original Article

Plugs clog the glandular outlets in fundic gland polyps

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Abstract: A systematic histologic analysis of 62 gastric fundic gland polyps (FGP) was carried out. All FGP (100%) showed foveolar cells with hypertrophic cytoplasm. In 95% of the FGP, parietal cells ballooned into the lumen and in 93%, exfoliated anucleated structures with eosinophilic granules were found. Plugs of anucleated structures with eosinophilic granules, most likely derived from exfoliated parietal cells, were found to clog the outlets of the glands in 86% of the FGP. None of the 30 control gastric biopsies without FGP had similar cellular aberrations. FGP seems to evolve by cellular aberrations affecting parietal cells. This is not surprising considering that genetic mutations are recorded in FGP with a common APC/b-catenin pathway in both FAP and sporadic cases. The genetic mutations in FGP might alter the biological behavior of the parietal cells, leading to increased exfoliation with clogging of the outlets of the glands. Thus, the blocking of the glandular outflow by plugs of anucleated structures with eosinophilic granules is the most likely cause for the cystic accumulation of “normal” glandular secretions.

Key words: Fundic gland polyps, eosinophilic plugs, cystic formations

Introduction

Fundic gland polyps (FGP) are common benign epithelial expansions of the fundic mucosa in patients having either hereditary diseases, such as familial adenomatous polyposis (FAP), Gartner’s syndrome, attenuated familial adenomatous polyposis syndromes, Peutz-Jeghers syndrome, Cowden’s syndrome, juvenile polyposis, or non-hereditary disorders (sporadic or somatic) such as Zollinger-Ellison syndrome, atrophic gastritis and medication with proton-pump inhibitors [1-23].

First described by Japanese researchers [24, 25], FGP are the most common gastric lesion in patients with FAP, with a reported prevalence between 27% and 73% [9, 10]. In the non-FAP population the prevalence of FGP is estimated to be between 0.8% and 1.4% [13, 21].

Lately, while reading gastric biopsies [26] having FGP, we found additional histological alterations to those previously reported in the literature.

The purpose of the present work was to describe and illustrate these not previously reported histological features in FGP and to assess their frequency in a cohort of patients with FAP and in patients without that genetic association.

Materials and methods

A total of 62 FGP removed at gastroscopy were analyzed: 33 were from FAP patients (hereditary FGP) and the remaining 29 were from patients without that genetic association (sporadic or somatic FGP). None of the patients received PPI medication. Fifteen sets of gastric biopsies from patients with Helicobacter pylori induced gastritis and 15 from patients with dyspeptic symptoms having normal gastric biopsies, were also reviewed.

Gastric biopsies were fixed in 4% neutral formalin; sections were stained with hematoxylin-eosin (H&E). Twenty of the 62 FGP sections were also stained with PAS.

All sections were scrutinized at high power microscopy (x40).

As FGP were regarded focal areas of the fundic mucosa with cystic dilatations lined with parietal, chief cells and occasional mucous foveolar cells.
A systematic histologic analysis of the FGP (Figure 1) was carried out starting from the top of the polyps as follows: the foveolar epitheli- um, the parietal cells subjacent to the foveolar epithelium, the outlets of glandular cysts and the cellular components wall-papering the fundic cysts.

Results

Foveolar epithelium: The cytoplasm of the foveolar cells were abnormally tall in all 100% (62/62) of the FGP. The hypertrophic cytoplasm contained excessive mucin, easily demonstrated with PAS stain (Figure 2).

Figure 2. Foveolar cells with hypertrophic cytoplasm containing excessive mucin (PAS x20).

Parietal cells subjacent to the foveolar epithelium: Near the foveolar epithelium the parietal cells were found to protrude into the lumen in 95.2% (59/62) (Figure 3). In addition, exfoliated anucleated cells with eosinophilic cytoplasmic granules, resembling the cytoplasm of parietal cells were found near the foveolar epithelium in 91.9% (57/62) of the FGP (Figure 4). No apoptotic granules were observed in these rounded eosinophilic structures.

Figure 3. Ballooning parietal cells near the foveolar epithelium (H&E x20).

Figure 4. Exfoliating anucleated structures with eosinophilic cytoplasmic granules, resembling the cytoplasm of parietal cells (H&E x20).
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Plugs in the outlets of the glandular cysts: Masses of anucleated, eosinophilic, granulated material (most likely derived from exfoliated parietal cells) or of more homogeneous stuff, were seen in the plugs that clogged the outlets of the fundic glands in 85.5% (53/62) of the FGP (Figures 5-7).

Plugs outside the outlets of the glandular cysts. Plugs outside the luminal aspect of the outlet of the glands (Figure 8) were found in 5 of the remaining 9 FGP (3 in FAP and 2 in sporadic cases. In addition, isolated eosinophilic clumps replacing the normal glandular epithelium were recorded in the remaining 4 FGP (3 in FAP case and 1 in a sporadic case.

Cellular components wall-papering fundic cysts: The parietal cells as well as other cells coating the cysts appeared flattened in the dilated fundic glands of all (100%) 62 FGP (Figure 1). The retained secretion seemed to have compressed the epithelium lining of the cysts.

No essential differences were found between the occurrence of the above-mentioned

Figure 7. Plug clogging the outlet of another gastric gland. The plug is built of anucleated, eosinophilic, granulated material, most likely derived from exfoliated parietal cells (H&E x20).

Figure 8. Plug, apparently outside the luminal aspect of the outlet of the glands. Note the anucleated, eosinophilic, granulated material in the plug, similar to the granulated cytoplasm in surrounding parietal cells (H&E x20).
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histological parameters in FAP cases and in sporadic cases.

None of the 30 control gastric biopsies without FGP, had similar cellular aberrations.

Discussion

The results of this investigation showed that in addition to the classically reported histological parameters characterizing FGP [1-25], several additional histological features were found such as the ballooning of the parietal cells, their abnormal exfoliation, the presence of anucleated, round structures with eosinophilic granules and the formation of plugs clogging the outlets of the glands.

The cystic formations in FGP would, therefore, evolve by the retention of the “normal” glandular secretions in obstructed glands. This obstruction appears to be the result of cellular alterations affecting parietal cells. This conclusion is based on the observation that parietal cells ballooned and subsequently exfoliated into the lumen of the glands. Once in the lumen, the exfoliated cells joined other exfoliated anucleated round formations with eosinophilic granules. These anucleated structures resembled the cytoplasm of parietal cells as no other cells in the area, had eosinophilic granules in their cytoplasm.

None of the 30 control gastric biopsies without FGP had similar cellular aberrations.

The finding of cellular aberrations in FGP is perhaps, not surprising considering that genetic mutations are often recorded in hereditary FGP (in the adenomatous polyposis coli gene (APC) and in sporadic FGP (in the b-catenin, a downstream target regulated by the APC protein) [27-36]. A common APC/b-catenin pathway seems to be involved in FGP through the targeting of different genes, both in FAP cases and in sporadic cases.

The gene ATP4A encodes the H+/K+ ATPase which is the major membrane constituent of parietal cells. Recently it was demonstrated that mice lacking H+/K+ ATPase (that is Atp4a(-/-) mice) developed countless gland cysts in the fundic mucosa [37]. Interestingly, the parietal cells in the knock-out Atp4a(-/-) mice are obviously affected, as these animals develop severe achlorhydria [37].

The claim that an obstructive mechanism is the cause for FGP is substantiated by the fact that a similar cell-obstructive mechanism was recorded in the glandular cysts often present in gastric adenomas [38]. The stratified dysplastic foveolar epithelium found at the luminal aspect of the gastric adenomas obstructed the glandular outlets of the normal gastric glands. Serial sections in gastric adenomas showed an inter-departmental communication between the labyrinths of cystically dilated glands [39]. The dysplastic epithelium initially detected at the luminal aspect of the adenomatous tissue, eventually replaced the entire epithelial layer of the cysts, in a downward growth fashion [38, 39]. More recently, we also found that the cause for the formation of retention cysts in colorectal adenomas was the obstruction of glandular outlets by dysplastic cells [40].

In the light of the present findings, several questions arise:

(i) If FGP is conveyed by a genetic aberration of the parietal cells, why not all cells in the fundic mucosa is also affected in patients with FGP? To answer that question appears to be as difficult as to answer why only some foveolar cells develop dysplatic changes leading to a gastric adenoma, while the rest of the foveolar epithelium appears normal, despite that all foveolar cells should have been subjected to the same carcinogen.

(ii) Why the parietal cells in FGP apparently develop regressive changes and detach from the gland resulting in abnormal exfoliation?

Further studies are necessary to unveil whether the genetic mutations in FGP alter the biological behavior of the parietal cells, leading to increased parietal cells exfoliation, resulting in the clogging of the outlets of the glands.

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