Silencing of TMSG1 enhances metastasis capacity by targeting V-ATPase in breast cancer

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Abstract: TMSG1, as a novel tumor metastasis suppressor gene, has been demonstrated to closely relate to the metastasis and drug-resistant of breast cancer. However, its molecular mechanism is still unclear. In this study, we explored the effect of small interference RNA (siRNA) targeting TMSG1 on the invasion of human breast carcinoma cell line MCF-7 and its molecular mechanisms associated with the extracellular pH. qRT-PCR and Western blot analysis revealed dramatic reduction of the levels of TMSG1 mRNA and protein after transfection of siRNA in MCF-7 cells. Cell migration and invasion were obviously increased by TMSG1 siRNA treatment. The activity of vacuolar ATPase (V-ATPase) and MMP-2 was significantly increased in MCF-7 cells transfected with the TMSG1 siRNA compared with the controls. Furthermore, acidic intracellular environment significantly increased the MMP-2 activity and the capacity of cell migration and invasion. In conclusion, silencing of TMSG1 increased V-ATPase activity, decreased extracellular pH and in turn the activation of secreted MMP-2, which ultimately promoted metastasis capacity of breast cancer cell.

Keywords: Breast cancer, TMSG1, V-ATPase, MMP-2, extracellular pH

Introduction

Breast cancer is the most common malignancy, threatening the health of women around the world. Although diagnostic and therapeutic methods have greatly improved over the last decade, more than 1.3 million women worldwide are diagnosed with breast cancer and about half-a-million women still die because of breast cancer each year [1, 2]. The high figures of incidences and mortality, even with the advancement of primary screening and diagnosis, increase the urgency to study the metastasis mechanism of breast cancer and explore the genes and treatment target associated with the metastasis of breast cancer.

Accumulating evidence suggests a key role of tumor acidic microenvironment in cancer development, progression, and metastasis [3-5]. V-ATPase, as a newly identified ATP-dependent proton pump, pumps H+ out of the cell membrane and thus play an important role in formation and maintenance of the extracellular acidic microenvironment [6-8]. V-ATPases are overexpressed in many types of metastatic cancers and positively correlated to their invasion and metastasis by decreasing extracellular pH and increasing secretion and activation of degradative enzymes, such as matrix metalloproteinases (MMPs) [6, 9]. Furthermore, low extracellular pH may promote the degradation and remodeling of ECM through proteolytic enzyme activation, thus contributing to cancer invasion and metastasis [10]. In breast cancer cells, the abundance of V-ATPase on the plasma membrane correlates with an invasive phenotype [11], and V-ATPase inhibitors reduce cell migration in cancer cells with high levels of plasma membrane V-ATPase [12, 13].

TMSG1 gene (tumor metastasis suppressor gene 1) is a novel tumor metastasis suppressor gene, which plays a prominent role in suppression of tumor invasion and metastasis [14]. It has been found that TMSG1 is negative correlation with tumor metastatic potential. Silencing of TMSG1 gene in PC-3M-2B4 cells enhanced invasion and metastasis of PCA cell [15]. The roles played by TMSG-1 in suppression of tumor
Silencing of TMSG1 enhances breast cancer cell metastasis capacity

invasion and metastasis have been reported to be achieved through various mechanisms. Recent studies showed that TMSG1 could regulate V-ATPase activity and extracellular pH by targeting the C subunit of V-ATPase, thereby induce apoptosis of tumor cells [16, 17]. In addition, TMSG1 also promotes cell apoptosis through the synthesis of ceramide, which in turn induces cell cycle arrest and affects the activity of telomerase as well as cell life span [18, 19]. Recently, it has been reported that low expression of LASS2 was associated with poor prognosis in patients with breast cancer [20]. However, the molecular mechanism by which TMSG1 on breast cancer metastasis is unclear at present.

In this study, small interfering RNAs (siRNAs) targeting TMSG1 will be adopted to silence the gene expression in human breast cancer cell line MCF-7. Meanwhile, we further elucidated the molecular mechanism of TMSG1 involved in the regulation of tumor metastasis by investigating the activity of V-ATPase, the expression and activity of matrix metalloproteinase 2 (MMP-2) in MCF-7 cells, then further revealed the key role of V-ATPase induced extracellular acidic microenvironment in metastasis of breast cancer.

Materials and methods

Cells and cell culture

MCF-7 breast cancer cells (American Type Culture Collection, Manassas, VA, USA; HTB-22) was cultured at 37°C in a humidified 5% CO₂ atmosphere in RPMI-1640 medium with 10% fetal calf serum (Gibco, Invitrogen, Carlsbad, CA, USA), 100 IU/ml penicillin G, and 100 mg/ml streptomycin sulfate (Sigma-Aldrich, St Louis, MO, USA).

Construction of siRNAs

We designed and purchased four different siRNA duplexes of TMSG1 from GenePharma company (Shanghai, China). siRNA-1 primers sense is 5’-CUGCCUUCUUUGGCUAUATT, and reverse is, 5’-UAUAUGGCAAAGAGGCAGTT-3’; siRNA-2 primers sense is 5’-GCUGCCUCUUGAACAUAATT, and reverse is, 5’-UUAUGUUCAGAGGGCAGTT-3’; siRNA-3 primers sense is 5’-GAGUCAGCAAAGUGUUAATT, and reverse is, 5’-UUAAACAUUCUUGGCGAGUCTT-3’; siRNA-negative control primers sense is 5’-UUCU-

CCGAACGUGUCACGUTT, and reverse is, 5’-ACGUGACACGUUCGGAGAATT-3’;

Transfection

Twenty-four hours before transfection cells were seeded, TMSG1 siRNA and Negative Control siRNA were transfected with Lipofectamine 2000 (Invitrogen, Carlsbad, CA) according to the manufacturer’s instructions. Twenty-four or forty-eight hours later, MCF-7 cells were collected and subjected to further analysis. Total RNA or protein was extracted from the indicated cells for analysis. The experiment was done in triplicates. More than nine wells were treated with the same kind of siRNA.

RNA extraction and qRT-PCR

Total RNA was isolated from cells with the TRizol reagent (Invitrogen) according to the manufacturer’s protocol. Real-time polymerase chain reaction was performed with the StepOne Plus sequence detection system (Applied Biosystems, Foster City, CA). To ensure the reproducibility of the results, all the genes were tested in triplicate. ITGB1 generated using sense (5’-gaagccagctggagattcac-3’) and antisense (5’-gacatcagaggcaatgctga-3’) primers. β-actin sense primers: 5’-cattaaggagaagctgtgct-3’ and antisense primers: 5’-gttgaaggtagttcgttggg-3’.

Western blot analysis

Whole cell extracts were prepared with a cell lysis reagent (Sigma-Aldrich, St. Louis, MO, USA) according to the manual, and then, the protein was quantified by a BCA assay (Pierce, Rockford, IL, USA). Then, the protein samples were separated by SDS-PAGE (10%) and detected by Western blot using polyclonal goat anti-TMSG1 antibody (Santa Cruz Bio-technology) or rabbit anti-MMP-2 antibody (Santa Cruz Bio-technology) and monoclonal mouse anti-β-actin (Santa Cruz Bio-technology) as a control. Goat anti-rabbit IgG (Pierce, Rockford, IL, USA) secondary antibody conjugated to horseradish peroxidase and ECL detection systems (SuperSignal West Femto, Pierce) were used for detection.

Activity of V-ATPase

Assays V-ATPase activity was performed as described previously by Fan et al [21].
Silencing of TMSG1 enhances breast cancer cell metastasis capacity

Gelatin zymography

MMP Zymography assay kit (for MMP-2 and MMP-9) (Applygen Technologie, China) was used to detect the activity of MMP2 and MMP9. Protein extracts and positive mixture were mixed with an equal volume of 2X SDS-PAGE non-reducing buffer, and electrophoresed on 8% SDS polyacrylamide gels containing 2 mg/ml of gelatin. Gels were then washed twice for 30 min in buffer A at room temperature, and incubated for 4 hours at room temperature in incubation buffer B. Gels were then stained for 2 hour with 0.25% Coomassie brilliant blue and then destained in destaining buffer (10% acetic acid and 20% methanol) for 60 min.

Invasion assay

Cells were cultivated to 80% confluence on the 12-well plates. Then, we observed the procedures of cellular growth at 24 h. All the experiments were repeated in triplicate. The transwell invasion chambers were used to evaluate cell invasion. Then cells invading across the membrane were counted under a light microscope.

Figure 1. TMSG1 siRNAs silenced the expression of TMSG1 in MCF-7 cells. A. qRT-PCR analyzed the expression of TMSG1 after transfection with different siRNA fragments in MCF-7 cells. B. The western blot analysis showed the expression of TMSG1 protein after interference. Control, untreated cells. siRNA NC, transfection with negative control siRNA. siRNA1, transfection with TMSG1 siRNA 1. siRNA2, transfection with TMSG1 siRNA 2. siRNA3, transfection with TMSG1 siRNA 3. Data are the mean ± SD of duplicates from a representative experiment of three independent experiments. *P < 0.01 vs. control and siRNA NC group.

Wound healing assay

For the wound healing assay, cells were seeded in 12-well plates and grown to 90% confluence. Monolayers in the center of the wells were scraped with pipette tips and washed with PBS. Cell movement into the wound area was monitored and photographed at 0 and 24 h using a light microscope. The migration distance between the leading edge of the migrating cells and the edge of the wound was compared as previous work [22].

Statistical analysis

Each experiment was repeated at least three times. Data were shown as mean ± s.d and analyzed using SPSS 18.0. Statistical comparisons between groups were analyzed using Student’s t-test and a two-tailed P < 0.05 was considered to indicate statistical significance.

Results

Effects of TMSG1 siRNAs on the expression of TMSG1 in MCF-7 cells

The MCF-7 cells were transfected with TMSG1 siRNAs and with negative control siRNA con-
Silencing of TMSG1 enhances breast cancer cell metastasis capacity

We examined four siRNAs to target human TMSG1 as described above. qRT-PCR analysis showed that siRNA-2 effectively blocked the expression of TMSG1 mRNA. qRT-PCR revealed dramatic reduction of 88.1% with siRNA-2, 60.1% with siRNA-3, and 12.2% with siRNA-1 in the levels of TMSG1 mRNA after transfection of siRNA in MCF-7 cells, compared with TMSG1 negative control siRNA or the control (untransfected) group ($P < 0.05$; Figure 1). Thus, the cells transfected with TMSG1 shRNA2 were used for further experiments.

**TMSG1 siRNA promotes the migration and invasion of MCF-7 cells**

As shown in Figure 2A, tranwell assays without Matrigel demonstrated that TMSG1 siRNA could significantly promoted migration of MCR-7 cells when compared with siRNA NC group and control (untreated) group. To further confirm this observation, we also determined the migration ability of breast cancer cells in the condition of TMSG1 silencing using a wound migration assay. The

Figure 2. TMSG1 siRNA affects the migration and invasion of MCF-7 cells. A. Transwell assays showed that the TMSG1 siRNA transfected cells displayed dramatically increased invasion ability compared with the untreated cells or Negative Control siRNA transfected cells. B. In wound migration assay, the cell-free wound gaps of MCF-7 cells monolayers healed slowly in the untreated cells or Negative Control siRNA transfected cells. Data are the mean ± SD of duplicates from a representative experiment of three independent experiments. *$P < 0.01$ vs. control and siRNA NC group.

Figure 3. Effects of TMSG1 siRNA on V-ATPase activity. Data are the mean ± SD of duplicates from a representative experiment of three independent experiments. *$P < 0.01$ vs. control and siRNA NC group.
results showed that the cell-free wound gaps of MCF-7 cells monolayers healed slowly in the untreated cells or Negative Control siRNA transfected cells. However, in TMSG1 siRNA-2 transfected cells, the closure of the wounded area was significantly accelerated (Figure 2B).

**TMSG1 siRNA increase the activity of V-ATPase in MCF-7 cells**

To explore whether TMSG1 siRNA suppress breast cancer cell migration and invasion by inhibiting the V-ATPase activity through binding to ATP6V1H, we examined the influence of TMSG1 siRNA on the V-ATPase activity by GEMEND’s V-ATPase activity assay kit. The activity of V-ATPase = OD of sample/(concentration × 31.1) × 10^3 U/mg. As shown in Figure 3, the V-ATPase activity was greatly increased in TMSG1 siRNA-2 treated MCF-7 cells (2.53 ± 0.031) compared with untreated cells (1.43 ± 0.022) and siRNA-NC treated cells (1.39 ± 0.014).
Effects of TMSG1 siRNA on the expression and activity of MMP-2 protein

V-ATPases pumps proton out of the cell membrane and thus plays an important role in formation and maintenance of the extracellular acidic microenvironment, which positively are positively correlated to secretion and activation of degradative enzymes, such as matrix metalloproteinases (MMPs) [6, 9]. Here, we determined the expression levels and activities of MMP-2, which are closed related to cancer invasion and metastasis [23]. Western blot showed that TMSG1 siRNA didn’t have obvious effect on the expression of total MMP-2 protein in the MCF-7 cells (Figure 4A). Furthermore, the supernatant of cultured cells was collected and the gelatinase activity was assayed with gelatin zymography. As shown in Figure 4B, the activity of MMP-2 was significantly increased by TMSG1 siRNA. These results indicated that TMSG1 didn’t have obvious effects on the expression levels of MMP-2 protein, but have significant effects on the activity of MMP-2 protein in the MCF-7 cells.

Extracellular pH affects the activity of MMP-2 protein and the migration and invasion of MCF-7 cells

Accumulating studies show a key role of tumor acidic microenvironment in cancer development, progression, and metastasis. The acidic tumor extracellular microenvironment induces the increased secretion and activation of proteases, and promotes the degradation and remodeling of extracellular matrix (ECM) through proteolytic enzyme activation, thus contribut-
Silencing of TMSG1 enhances breast cancer cell metastasis capacity

...ing to cancer invasion and metastasis. In this study, we further analyzed the effects of acidic extracellular microenvironment on MMP-2 protein in MCF-7 cells. As shown in Figure 5A, extracellular pH didn’t affect obviously the expression of total MMP-2 protein in the MCF-7 cells using western blot analysis. Furthermore, the gelatinase activity assayed the revealed the positive correlation between extracellular pH and MMP-2 activity. As shown in Figure 5B, the activity of MMP-2 was significantly increased in a PH-dependent manner. These results indicated that extracellular pH didn’t have obvious effects on the expression levels of MMP-2 protein, but have significant effects on the activity of MMP-2 protein in the MCF-7 cells.

Furthermore, we demonstrated the effects of extracellular PH on invasion and migration of MCF-7 cells. As shown in Figure 6A, acidic extracellular microenvironment (PH 6.6) dramatically increased invasion ability compared with the PH 7.0 and PH 7.4 extracellular microenvironment. The wound migration assay further confirm this observation that acidic extracellular microenvironment greatly increased migration ability compared with the PH 7.0 and PH 7.4 extracellular microenvironment (Figure 6B).

Therefore, acidic extracellular microenvironment significantly increased the migration and invasion ability of MCF-7 cells.

Discussion

Tumor metastasis suppressor gene 1 (TMSG1), also known as LASS2, is a newly found tumor metastasis suppressor gene. Accumulating evidences showed that it not only suppressed tumor growth but was closely related to tumor metastasis. Recently, it has been reported that expression levels of TMSG-1 was altered in breast cancer and TMSG-1 overexpression inhibited the proliferation, invasion and apoptosis of metastatic breast cancer cells significantly [20, 24]. However, as a novel TMSG, the precise molecular mechanisms of TMSG1 on breast cancer metastasis are unclear.

Therefore, to further investigate the molecular mechanisms of the antitumor effect of TMSG-1 on metastasis of breast cancer, small interference RNA (siRNA) was adopted to silence the gene expression of TMSG1 in human breast cancer cell line MCF-7. Furthermore, we confirmed that siRNA2 could significantly decreased the TMSG1 expression at the mRNA level and protein level, reduced 88.1% TMSG1 mRNA and 93.9% TMSG-1 protein. Then, we further analyzed the effects of LASS2 silencing on invasion and migration of breast cancer. Our results showed that silencing of TMSG1 could promote the migration and invasion of MCF-7 cells, which in turn suggested that TMSG1 gene function as tumor metastasis suppressor.

It is well-known that extracellular pH is usually low in solid tumors, in contrast to the approximately neutral intracellular pH. V-ATPase, as ATP dependent hydrogen ion pump, pumps out H+ through transmembrane transportation and plays important roles in formation and maintenance of extracellular acidic microenvironment and metastasis [9]. Recently, Sennoune et al. showed the correlation of V-ATPases located at the plasma membrane with the metastatic potential of breast cancer cells [12]. The preferential expression of V-ATPase at the cell surface is important for the acquisition of invasiveness and metastasis of the breast cancer cells [12]. Moreover, TMSG1, as a suppressor of V-ATPase, inhibited the V-ATPase activity by interacting with ATP6V0C, the c subunit of V-ATPase proton pump [16]. Here, in order to investigate the breast tumor metastasis suppression mechanism of TMSG1, we studied the interaction between TMSG1 and V-ATPase. As shown in Figure 3, the activity of V-ATPase was greatly increased byTMSG1 siRNA in MCF-7 cells. The findings were also confirmed in previous work that TMSG1 siRNA and shRNA could enhance the activity of V-ATPase and increase extracellular H+ concentration in prostate cancer cells [15, 25]. These results suggested that TMSG-1 could inhibit the activity of V-ATPase, suppress the proton transmembrane secretion and thus decrease extracellular H+ concentration in breast cancer cells. The promoting effect of V-ATPase on cancer invasion and metastasis mainly relies on its maintaining acidic pH of extracellular microenvironment, which is related to the activation, secretion, and cellular distribution of many proteases involved in the digestion of ECM [11]. The pH-sensitive proteases include cathepsin and MMPs (MMP-2, MMP-9, MMP-3, etc.). Here, we demonstrated that TMSG1 siRNA could increase the activity of...
Silencing of TMSG1 enhances breast cancer cell metastasis capacity

MMP-2 in the supernatant of the MCF-7 cells, but had no effect on the expression and secretion of MMP-2 protein, which was consistent with previous studies by Xu et al [15, 25]. These results indicated that migration of tumor cells might be regulated by MMP activity, rather than by MMP expression.

To reveal the relationship among V-ATPase-induced acidic extracellular microenvironment, tumor metastasis and MMP2 protein, we further analyzed invasion and migration capacity of MCF-7 cells at PH 6.6, PH 7.0 and PH 7.4 extracellular microenvironment. As shown in Figure 5, MMP2 expression level in MCF-7 cells was not affected by different PH cultured conditions, however, the MMP2 activity was significantly upregulated from pH 7.4 and pH 6.6 cultured conditions. Furthermore, acidic extracellular microenvironment (PH 6.6) dramatically increased invasion and migration ability compared with the PH 7.0 and PH 7.4 extracellular microenvironment. TGMS1 silencing- induced invasion and migration were reversed by high extracellular pH (PH 7.4). These data also indicated that V-ATPase played an important role in breast cancer metastasis.

In conclusion, silencing of TMSG1 promoted the formation of the extracellular acidic microenvironment by targeting V-ATPase and promoting the V-ATPase activity in breast cancer. The acidic extracellular microenvironment activated degradative enzymes, such as matrix metalloproteinases (MMPs) and degradation of ECM, ultimately accelerated tumor’s invasion and metastasis. Our findings suggest that TGMS1, as a novel tumor metastasis suppressor gene, is a promising therapeutic target for advanced breast cancer.

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Disclosure of conflict of interest

None.

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Silencing of TMSG1 enhances breast cancer cell metastasis capacity


