Original Article
Expression and correlation of matrix metalloproteinase-7 and interleukin-15 in human osteoarthritis

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Abstract: Objectives: To investigate the expression and correlation of matrix metalloproteinase (MMP)-7 and interleukin (IL)-15 in human osteoarthritis (OA). Methods: From October 2013 to December 2014, 30 patients with OA were enrolled. In addition, another 30 patients with simple meniscus injury were collected as a control group. There were no significant differences in age and gender between the two groups. Articular cartilage tissue was obtained from both OA patients and control group patients. Protein, mRNA, and serum expression levels of MMP-7 and IL-15 in the two groups were determined by immunohistochemical (IHC), in situ hybridization, and enzyme-linked immunosorbent assay (ELISA) assay, respectively. Additionally, correlation between MMP-7 and IL-15 expression level in cartilage tissue and serum was assessed using Pearson correlation analysis. Results: Protein, mRNA, and serum expression levels of MMP-7 and IL-15 in patients with OA were all significantly increased in OA patients compared with the control group. Besides, there were strong positive relationships between articular MMP-7 level and serum MMP-7 level ($R^2 = 0.573, P = 0.018$), between articular IL-15 level and serum IL-15 level ($R^2 = 0.861, P = 0.023$), and between serum IL-15 level and serum MMP-7 level ($R^2 = 0.602, P = 0.012$). Conclusion: These results suggest that MMP-7 and IL-15 might play important roles in the pathogenesis of OA, and IL-15 and MMP-7 has positive correlation in OA.

Keywords: Matrix metalloproteinase-7, interleukin-15, osteoarthritis

Introduction

Osteoarthritis (OA) is the most common joint disorder, which particularly affects the weight-bearing joints, predominantly in the knee [1]. It is characterized by progressive loss of cartilage matrix, subchondral bone remodeling, and osteophyte formation, etc. [2, 3]. It has been reported that approximately 10% people over age 60 years in the world suffer from OA [2], and the prevalence is estimated to continue to grow [4]. Besides, OA not only impacts on an individual’s health, but also carries out a significantly financial and societal burden for families and the society [3, 5]. However, effective management of OA has not been acquired due to the unclear pathophysiology mechanism. Recently, roles of matrix metalloproteinase (MMPs) and cytokines in OA have been increasingly paid attention. MMPs and cytokines are considered as potential candidates for biochemical mediators [6]. The mediator either acted individually, or with another mediator acted in networks could greatly regulate cellular responses (both catabolic and anabolic activities) in joint tissues [7]. MMPs are a family of Zn$^{2+}$-dependent endopeptidases that participate in the degradation of extracellular matrix (ECM). Previous studies have indicated that MMP-1, -2, -3, -8, -9, and -13 are involved in OA [8-11], and MMP-13 has been regarded as a crucial target gene for the progression of OA [12]. Besides, a study conducted by Ohta suggested that MMP-7 is overproduced in human
OA cartilage and MMP-7 may play a significant role in the ECM degradation in OA [13]. Cytokines, such as interleukin (IL)-1, IL-6, IL-15 and tumor necrosis factor (TNF)-α, have been involved in the pathogenic mechanism of OA [14-17]. Among the cytokines, IL-15 has been regarded as a candidate therapeutic target for OA [17]. However, little information is available regarding the associations of MMP-7 expression and IL-15 expression in articular and serum of OA patients.

Therefore, we aimed to investigate the MMP-7 expression and IL-15 expression in both articular and serum of OA patients, as well as the associations between articular and serum of MMP-7, the associations between articular and serum of IL-15 level, and the associations between serum of MMP-7 expression and IL-15 expression. Our study might contribute to explore the pathogenesis mechanism of OA.

**Materials and methods**

*Participants and study design*

This study was approved by the hospital medical ethics committees, and informed consent was obtained from all participants. Between October 2013 and December 2014, thirty patients with knee OA (13 males and 17 females; mean age 65.16 ± 17.66, range 35-76) attending at our hospital were enrolled. Besides, thirty patients with simple meniscus injury (16 males and 14 females; mean age 63.60 ± 15.23, range 30-79) were collected as a control group. Diagnosis of OA was based on clinical and radiological evaluations revealed by the 1986 American College of Rheumatology (ACR) criteria [18]. Patients with obvious joint injury or with inflammatoy arthritis, previous knee surgery were excluded from the study. Also, patients who received intra-articular corti-
costeroid or hyaluronan injections within 3 months of surgery were excluded.

Cartilage tissue samples

Articular cartilage was acquired from both OA patients and meniscus injury patients who were undergoing arthroscopic examination or arthroplastic surgery. The tissue sample was fixed in 4% paraformaldehyde immediately for 24 h after biopsy. Then the tissue was embedded in paraffin, sectioned, and dewaxed according to the standard methods [13]. Consecutive 4-μm-thickness sections were prepared.

Immunohistochemical (IHC) staining

IHC staining was performed using monoclonal anti-MMP-7 antibody (diluted 1:50; Chemicon, Temecula, CA) and anti-IL-15 antibody (diluted 1:200; Santa Cruz Biotechnologies). The slides were pretreated by microwave heating before the incubation. After incubation with primary antibody, biotinylated secondary antibody was performed. Negative controls procedures were also performed (the primary antibody was replaced by phosphate-buffered saline (PBS) solution). Finally, chromogen 3,3-diaminobenzidine-hydrogen peroxide (DAB, Sigma-Alderich, S.R.L) was performed for visualized reaction.

In situ hybridization

In situ hybridization was performed on 4-μm-thickness sections according to a standard method [19] with a MMP-7 kit (HPV-0610, Fuzhou Maixin Biotech. Co. Ltd) and a IL-15 kit (MK1165, Boster, Wuhan, China). Briefly, the sections were deparaffinized, treated with 0.01% Triton X-100 for 2 min, deproteinated with proteinase K (0.25 g/L) for 5 min and washed in 2 × saline sodium citrate (SSC) at 42°C. Then the slides were prehybridized at 42°C for 3-4 h in prehybridization solution. The probe of MMP-7 and IL-15, and 20 pg tRNA were mixed with the prehybridization solution.
Expression and correlation of MMP-7 and IL-15

and incubated at 42°C overnight, and washed with 2 × SSC. Thereafter, 0.1% Triton X-100 was performed for 2 min and washed with buffer solution 1 (1 mol/L NACl, 0.1 mol/L Tris, PH7.5). The slides were then incubated with buffer solution 2 (buffer solution 1 and 3 g bovine serum albumin (BSA)) for 1 h. After the buffer solution 2 was discarded, avidin and alkaline phosphatase was added to the mixture for 20 min at 42°C. Finally, autoradiography was carried out by nitroblue tetrazolium (NBT).

Computer-assisted image analysis

Five representative areas of each sample (assessed by both IHC and in situ hybridization) were randomly chosen under a light microscope (Olympus, Tokyo, Japan). Quantitative analysis was performed with Image-Pro Plus 6.0 (Media Cybernetics, GE, USA). The positive areas were regarded as specific brown yellow color (for IHC) or blueviolet (for in situ hybridization). The integrated optical density (IOD; pixel area × the optical density) was measured after standard optical density (OD) calibration.

Enzyme-linked immunosorbent assay (ELISA) assay

Fasting blood samples (5 mL) were obtained from an antecubital vein. After clotting, the blood samples were separated by centrifugation at 1,000 g for 15 min at 4°C. Serum samples were subsequently stored at -80°C until use. The serum levels of MMP-7 and IL-15 were determined by an ELISA kit (Calbiochem, Cambridge, MA, USA) according to the manufacturer’s protocol. Briefly, 96-well plates were coated with either anti-MMP-7 or anti-IL-15 antibody, and the appropriate diluent serum samples were added in duplicate, followed with incubation for 2 h at room temperature. After washing with wash buffer, the corresponding secondary antibody was applied. After incubation for another 2 h, the plates were washed again. Spectrophotometric microplate reader was employed to determine the optical density at 490 nm. The concentration of MMP-7 and IL-15 was calculated from a standard curve.

Statistical analysis

Continuous variables were expressed as the mean ± standard deviation (SD). Statistical analysis was performed with statistical package for the social sciences (SPSS) software (version 17.0; SPSS Inc., Chicago, IL). The 2-sample Student’s t test was used to compare continuous variables. Correlation analysis was performed using Pearson test. A statistical significance was defined when \( P < 0.05 \).

Results

MMP-7 and IL-15 protein and mRNA expression

IHC and in situ hybridization were performed to examine the protein and mRNA expression levels of MMP-7 and IL-15 in human cartilage tissue, respectively. Here we set the baseline of relative IOD in control group to be 1. The results showed that the expression protein levels of MMP-7 and IL-15 were both significantly increased in OA patients compared with control group (\( P < 0.05 \)) (Figure 1A and 1B). Similarly, the results of in situ hybridization demonstrated that the expression mRNA levels of MMP-7 and IL-15 were also both statistically increased in patients with OA compared with the control group (\( P < 0.05 \)) (Figure 2A and 2B).

MMP-7 and IL-15 serum expression

The serum expression levels of MMP-7 and IL-15 were showed in Figure 3A and 3B, respectively. As shown in Figure 3A, the average serum MMP-7 level was 5.617 ± 2.300 ng/ml (range 2.2-8.8 ng/ml) in the control group, whereas the average serum MMP-7 level was 20.117 ± 7.083 ng/ml (range 11.0-29.4 ng/ml). There were significant differences in the serum MMP-7 level between the two groups. Also, the serum IL-15 level (average 129.6 ± 39.922 pg/ml; range 85.0-176.0 pg/ml) in OA patients was significantly higher than that in the control group (average 576.8 ± 194.574 pg/ml; range 369.0-875.0 pg/ml) (Figure 3B).

Correlation analyses between MMP-7 and IL-15

In order to confirm the clinical diagnostic value of MMP-7 and IL-15 in OA, we further performed the correlation analysis between the expression level of serum and articular cartilage specimens in OA patients. The results showed that there were strong relationships between articular MMP-7 level and serum MMP-7 level (\( R^2 = 0.573, P = 0.018 \)), between articular IL-15 level and serum IL-15 level (\( R^2 = 0.861, P = 0.023 \)), and between serum IL-15 level and serum
MMP-7 level ($R^2 = 0.602, P = 0.012$) (Figure 4A-C).

Discussion

In the present study, the expression and correlation of MMP-7 and IL-15 in human OA was investigated. The results showed that the protein, mRNA, and serum expression levels of MMP-7 and IL-15 in OA patients were all significantly higher than those in the control group. Besides, there were strong positive relationships between articular MMP-7 level and serum MMP-7 level, between articular IL-15 level and serum IL-15 level, and between serum IL-15 level and serum MMP-7 level. Our results indicated that MMP-7 and IL-15 might play important roles in the pathogenesis of OA, and IL-15 and MMP-7 has positive correlation in OA.

MMPs are a group of proteolytic enzymes responsible for the degradation of ECM, which is involved in multiple physiological and pathological progressions of different tissues [20-22]. The degradation of ECM plays essential roles in the tissue resorption and remodeling [23]. It has been reported that approximately 95% of the dry weight of articular cartilage is ECM [24]. The imbalance of the cartilage ECM results in OA, therefore, a delicate balance between the breakdown and synthesis of ECM is of importance [25]. MMP-7, also known as matrilysin, is a unique member of the MMP family. Unlike other members, MMP-7 is the smallest member that only consists of the common catalytic domain and zinc-binding region [26]. It has a specific ability to degrade various ECM components such as cartilage proteoglycan [27]. Since Ohta has firstly reported that MMP-7 is overexpressed in human OA cartilage [13], the functions and roles of MMP-7 in OA have been paid attention [28, 29]. In consistent with previous studies, our results also suggested that the expression levels of MMP-7 were significantly higher than those in the control group, either in articular cartilage or in serum. Additionally, we found that there was a strong positive relationship between articular MMP-7 level and serum MMP-7 level.

The expression of MMPs is regulated by cytokines [30]. Inflammatory reaction has been well demonstrated in the development and progression of OA, even in the early stage of the disease [31]. IL-15, a proinflammatory cytokine, has been considered as a contributor to inflammation in OA [6]. Previous study has indicated that IL-15 has both potential diagnostic and prognostic biochemical marker for OA [28]. The expression level of IL-15 was found significantly increased in OA patient’s serum samples; besides, IL-15 level was positively correlated with a Western Ontario McMaster University Osteoarthritis Index (WOMAC) pain scores [32]. Moreover, IL-15 has been reported to induce MMP production, especially MMP-1 and MMP-9 [33]. Additionally, previous studies suggested that there were associations between IL-15 level with MMP-1 and MMP-3 [17]. Also, our study confirmed the higher expression of IL-15 in OA patients than that in control patients, which was in line with previous studies. In addition to the results, we found that serum IL-15 level was positively related with articular IL-15 level and serum MMP-7 level. One possible reason for that is MMP-7 is may be induced by IL-15, and both of them participate in the pathogenesis of OA.

In conclusion, our results suggest that both MMP-7 and IL-15 play important roles in the pathogenesis of OA, and IL-15 and MMP-7 has positive correlation in OA.

Disclosure of conflict of interest

None.

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Expression and correlation of MMP-7 and IL-15


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Expression and correlation of MMP-7 and IL-15


