Original Article

Implication of Reprimo and hMLH1 gene methylation in early diagnosis of gastric carcinoma

Lianhua Liu¹, Xiaofeng Yang²

¹Department of Radiotherapy, Lin-Yi Cancer Hospital, Linyi 276001, Shandong, China; ²Department of Emergency Surgery, Lin-Yi People’s Hospital, Linyi 276001, Shandong, China

Received September 6, 2015; Accepted October 21, 2015; Epub November 1, 2015; Published November 15, 2015

Abstract: DNA methylation has been recently recognized as a novel tumor marker. This study investigated the methylation status of Reprimo and hMLH1 gene in both plasma and tissue samples from gastric cancer patients, in an attempt to investigate their diagnostic implications in gastric cancer. A total of 180 tissue and plasma samples (including 50 cases of gastric cancer, 50 dysplasia, 50 chronic atrophic gastritis with intestinal metaplasia and 30 normal controls) were collected for detecting DNA methylation status of Reprimo and hMLH1 genes using MSP method. Tissue protein expression levels were further tested by immunohistochemical (IHC) staining. The positive rate of DNA methylation rate was, in ascending sequence, gastritis tissue, dysplasia tissue and gastric carcinoma tissue. All those tissues had significantly elevated DNA methylation level compared to normal group (P < 0.05). Expression level of Reprimo and hMLH1 proteins were, however, decreased in pathological tissues compared to normal ones (P < 0.05). A significantly negative relationship existed between protein level and promoter region methylation level. The DNA methylation occurred in promoter regions of both Reprimo and hMLH1 genes depressed the protein expression, and may participate in the occurrence and progression and gastric cancer. The combined assay of serum Reprimo and hMLH1 DNA methylation levels thus had critical importance in the early diagnosis and gastric cancer.

Keywords: DNA methylation, reprimo gene, hMLH1 gene, gastric cancer, early tumor diagnosis

Introduction

As one common malignant tumor in digestive tract, the pathogenesis of gastric cancer involves a continuously pathogenic process from chronic gastritis towards mucosal atrophy, intestinal metaplasia, dysplasia and eventually tumors [1]. The underlying genetic alternation thus owns critical values for the early diagnosis and exploration of pathogenesis.

Besides genetic mutation, epigenetic alternation is also correlated with gastric cancer. DNA methylation is the most common epigenetic mechanism, involving the addition of methyl group to 5'-carbon of cytosine under the direction of DNA transferase [2]. DNA methylation may affect the expression of tumor suppressor gene and mismatch repair gene, both of which participate in processes including cell cycle modulation, DNA repair, oncogenic metabolites, intracellular interaction, apoptosis and angiogenesis [3]. Specifically, the methylation of CpG island in promoter region of tumor suppressor gene and mismatch repair gene may impede the binding between promoter and transcriptional factors, thus promoting the occurrence and progression of tumors [4]. Studies have revealed the existence of CpG island hyper-methylation in almost all human tumor tissues, especially during the early stage [5]. Such tumor-specific DNA methylation can be detected in blood samples, suggesting its potency as one non-invasive tumor marker [6, 7].

Located on human chromosome 2, Reprimo gene is one of genes that regulating normal cell growth. The deactivation of Reprimo gene occurs at the early stage of multiple tumors, mainly due to DNA methylation [8]. In one study of gastric cancer patients’ plasma and tissue samples, the positive rate of Reprimo gene methylation was 95.3% and 97.7% respectively, in sharp contrast to 9.7% in normal people [9]. As the major component of DNA mis-
match repair system, hMLH1 gene is also correlated with gastric cancer pathogenesis. In most tumor-adjacent tissues from gastric cancer patients, methylation can be found in gene promoter region, suggesting that hMLH1 gene methylation may be one important molecular event during the early progression of gastric cancer [10, 11].

Currently no study has been performed regarding the abnormal expression of both tumor suppressor gene Reprimo and mismatch repair gene hMLH1 in various stages during the occurrence of gastric cancer. This study thus for the first time tested the expression of Reprimo and hMLH1 gene in both peripheral blood and tissue samples from gastric cancer patients, in an attempt to analyze the correlation between those two genes and different stages of gastric cancer, thus providing novel molecular marker for the early diagnosis of gastric carcinoma.

Materials and methods

Study objects

A total of 180 individuals were recruited in the Lin-Yi Cancer Hospital from October 2012 to October 2014. Tissue samples were collected from gastroscopy. Inclusive criteria were: (1) Confirmed pathological diagnosis as chronic atrophy gastritis complicated with intestinal metaplasia, dysplasia or gastric carcinoma; (2) All participants have signed written consents with fully understanding of this study. Exclusive criteria: (1) Complicated with other digestive diseases; (2) With other systemic tumors and/or metastasis to the gastric; (3) Have undergone surgical or chemotherapy; (4) Complicated with other severe systematic disease or organ failure. A total of 50 chronic atrophy gastritis, 50 cases of dysplasia, 50 cases of gastric carcinoma and 30 normal gastric mucosal tissues. The general information of all people involved is listed in Table 1. No significant difference regarding age or sex distribution has been discovered between control and diseases animals.

DNA extraction and bisulfite modification

Fasting blood samples (5 mL) were collected from veins of all participants and were stored in EDTA-containing tubes. After centrifugation at 1 500 g for 10 min, plasma was saved to extract genomic DNA using DNA purification kit (Zymo Research, US). The purity of DNA was determined by agarose gel electrophoresis and UV spectrometer. After modification using bisulfite, DNA samples were kept at -20°C for further use.

MSP assay of plasma sample

Specific methylation (M) and un-methylation (U) primers were designed based on specific sequences of CpG islands of hMLH1 and Reprimo gene promoter regions [9, 10]. Sequences were: Reprimo-M-Forward, 5’-GCGAG TGAGC GTTTA GTTC-3’, Reverse, 5’-TACCT AA- AAC CGAA TTCAT CG-3’; Peprimo-U-Forward, 5’-TTGTG AGTGA GTGTT TAGTT TG-3’; Reverse, 5’-TAATT ACCTA AAACCA AATTCA TC-3; hMLH-1-M-Forward, 5’-TTGTG TGGAT ATTTT GTATT TTTTT G-3’; Reverse, 5’-CTCCCT AAAAC AACTA CTACCC; hMLH1-U forward, 5’-ATTGG TTGGAT ATTTT CGTA TTTTT C-3’; hMLH1-U reverse, 5’-CCTAA AACGA CTCTA CCCGC T-3’. PCR was carried under the following condition: 95°C for 5 min, followed by 30 cycles each containing 95°C denature for 30 sec, 62°C annealing for 60 sec and 72°C elongation for 30 sec. PCR products with methylation activity was employed as a positive control using healthy adjacent tissues without modification. Another cohort of positive control will be used for non-methylation assay. A gel imaging equipment (BIO-RAD, US) was used to capture pictures. The methylation status (Band M: methylated, DNA Bank U: un-methylated DNA).

Protein expression level

All tissue samples were immediately fixed in 10% neutral buffered formaldehyde (NBP). After embedding in paraffin, 4 μm-thickness consecutive slices were made. Protein expression levels of Reprimo and hMLH1 gene. A total of

Table 1. General information of all patients

<table>
<thead>
<tr>
<th>General info</th>
<th>Intestinal metaplasia</th>
<th>Dysplasia</th>
<th>Gastric cancer</th>
<th>Normal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>30</td>
<td>27</td>
<td>331</td>
<td>17</td>
</tr>
<tr>
<td>Female</td>
<td>20</td>
<td>23</td>
<td>17</td>
<td>12</td>
</tr>
<tr>
<td>Age</td>
<td>55.4±11.2</td>
<td>59.4±13.9</td>
<td>63.9±16.5</td>
<td>58.4±14.7</td>
</tr>
<tr>
<td>Range</td>
<td>32~69</td>
<td>37~73</td>
<td>42~79</td>
<td>35~78</td>
</tr>
</tbody>
</table>
Gene methylation in gastric cancer

12 high magnification (× 400) fields were randomly selected from the family. The number of IHC-positive cells was counted to define positive slices (i.e. those with less than 5% of NBF were regarded as negative slices and vice versa).

Statistical analysis

SPSS 16.0 software package was used to process all collected data, of which measurement data was presented as mean ± standard deviation (SD). Student t-test was used to compare means between two groups. Multiple group comparison was done by analysis of variance (ANOVA). Enumeration data, on the other hand, was tested by chi-square approach. A statistical significance was defined when \( P < 0.05 \).

Results

MSP amplification of plasma sample

Methylation assay of Reprimo gene showed that, 14 out of 50 samples with chronic atrophy gastritis with intestinal metaplasia showed gene methylation (positive rate = 26%). In all dysplasia samples, 28 cases had methylation (positive rate = 56%). In gastric cancer patients, this number was even as higher as 62% (31 out of 50). All 30 normal controlled people, however, did not have detectable methylation of Reprimo gene (Figure 1). Compared to those in control people, the methylation rates in intestinal metaplasia, dysplasia and gastric carcinoma patients were significantly elevated (\( P < 0.05 \)). Among those three groups, gastric cancer and dysplasia had highest level of methylation, as compared to intestinal metaplasia and control people (\( P < 0.05 \)).

Methylation assay for hMLH1 gene showed that, 10 out of 50 in gastritis with intestinal metaplasia, 22 out of 50 of dysplasia patients, and 24 out of 50 gastric cancer patients had DNA methylation, making the positive rates at 20%, 44% and 48%, respectively. In contrast, healthy people had the methylation rate only 3.3% (1 out of 30 people, Figure 1). These data suggested that dysplasia and gastric cancer were accompanied with significantly higher hMLH1 gene methylation level compared to that in intestinal metaplasia patients (\( P < 0.05 \)), who further had higher methylation level than that in control group (\( P < 0.05 \)).

Protein expression level of Reprimo and hMLH1 genes

We further used IHC staining to visualize the protein expression level of Reprimo and hMLH1 genes in tissue samples. As shown in Figure 2, the positive expression rate of Reprimo and hMLH1 genes in intestinal metaplasia, dysplasia, gastric cancer and normal group were 20% (10/50) and 22% (11/50), 44% (22/50) and 40% (20/50), 48% (24/50) and 46% (23/50), and 3.3% (1/30) and 0%, respectively. Therefore, the positive expression rate of both genes were positively correlated with methylation rate in plasma (\( r = 0.99, P < 0.05 \)).

Combined assay of methylation of two genes

As shown in Table 2, the positive rates of combined methylation assay in the plasma of intestinal metaplasia, dysplasia, gastric cancer and normal group were 34%, 76%, 84% and 3.3%, respectively. The combined negative rates in tissue samples were 36%, 78%, 82% and 0%. Therefore, the combined assay of two gene markers can improve the specificity and sensitivity of diagnosis.

Discussion

The 5-year survival rate of early-stage gastric cancer can reach up to 95%. However, once the tumor has invaded into muscular or serous layer, the survival rate has dropped to less than 20% [12]. Therefore, the early intervention of gastric cancer can total cure the disease. However, due to the lack of unique symptom, the diagnostic rate of gastric cancer at its early
stage was less than 10%. Most patients were already at the advanced stage at the time of first diagnosis, leading to lower survival rate [13]. The effective early diagnostic approach is thus of significant values and has drawn lots of research interests [14-16]. The precancerous lesion of gastric carcinoma includes atrophy gastritis, intestinal metaplasia and intraepithelial neoplasia, all of which are risk factors for gastric cancer and can be used to improve the diagnostic rate at early stage for improving the prognosis. Recently, epigenetic studies including DNA methylation has provided more insights regarding the early diagnosis and prognostic prediction of tumors [17]. The methylation of both tumor suppressor gene and mismatch repair gene at their promoter region is closely correlated with tumor occurrence [18].

This study investigated the methylation status of tumor suppressor gene Reprimo and mismatch repair gene hMLH1 at their promoter regions, for further exploration of the potency as biological indexes for screening and early diagnosis of gastric cancer. As one newly discovered tumor suppressor gene, Reprimo codes a highly glycosylated protein in the cytoplasm for mediating p53-directed cell cycle. It can arrest abnormal cells at G2 phase, inhibit the Cdc2 activity and nuclear translocation of cyclin B1, and prevent hyper-proliferation of cells [19]. Studies have found that the deactivation of Reprimo gene occurred at the early stage of dozens of malignant tumors, mainly as a result of DNA methylation. For example, Reprimo gene methylation has been reported in most of gastric cancer, lymphoma and colorectal carcinoma, in contrast to minimal level in normal tissues [20, 21]. The abnormal methylation of Reprimo gene is thus a hallmark at the early stage of certain cancers. Recently, the correlation between mismatch repair system and gastric cancer occurrence has also been investigated [22, 23]. The major function of mismatch repair gene is to correct the mismatching of base pairs occurred during DNA

<table>
<thead>
<tr>
<th>Group</th>
<th>N</th>
<th>Reprimo methylation rate</th>
<th>Reprimo protein expression</th>
<th>hMLH1 methylation rate</th>
<th>hMLH1 protein expression</th>
<th>Combined rate of methylation</th>
<th>Combined expression rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intestinal metaplasia</td>
<td>50</td>
<td>14 (28%)</td>
<td>16 (32%)</td>
<td>10 (20%)</td>
<td>11 (22%)</td>
<td>17 (34%)</td>
<td>18 (36%)</td>
</tr>
<tr>
<td>Dysplasia</td>
<td>50</td>
<td>28 (56%)</td>
<td>30 (60%)</td>
<td>22 (44%)</td>
<td>20 (40%)</td>
<td>38 (76%)</td>
<td>39 (78%)</td>
</tr>
<tr>
<td>Gastric cancer</td>
<td>50</td>
<td>31 (62%)</td>
<td>33 (66%)</td>
<td>24 (48%)</td>
<td>23 (46%)</td>
<td>42 (84%)</td>
<td>41 (82%)</td>
</tr>
<tr>
<td>Normal</td>
<td>30</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
<td>1 (3.3%)</td>
<td>0 (0%)</td>
<td>1 (3.3%)</td>
<td>0 (0%)</td>
</tr>
</tbody>
</table>

Figure 2. Reprimo and hMLH1 protein expression (× 100). A-D. Reprimo; E-H. hMLH1 protein. A and E. Intestinal metaplasia; B and F. Dysplasia; C and G. Gastric cancer; D and H. Control.
Gene methylation in gastric cancer

replication, thus maintaining genome stability [24]. As one DNA mismatch repair gene, hMLH1 can affect the endogenous repair function of cells and has been suggested to play an important role in the occurrence of gastric cancer [25]. The microsatellite instability (MSI) is related with mismatch repair gene, and is one feature of gastric cancer [26]. Many studies have showing the deactivation of hMLH1 gene due to its hyper-methylation under MSI environment. The absence of MSI, however, helps to depress gene methylation level for facilitating gene expression, suggesting the possible involvement of hyper-methylation of hMLH1 gene promoter in gastric cancer [27].

To investigate the correlation between methylation of tumor suppressor gene Reprimo and mismatch repair gene hMLH1 and the occurrence of gastric cancer, we recruited patients including intestinal metaplasia, dysplasia and gastric cancer at different stages. On those patients, both MSP and IHC methods were used to detect the promoter methylation level and protein expression level in plasma and tissue samples. Our results showed elevated gene methylation level in intestinal metaplasia, dysplasia and gastric cancer patients, with an ascending order. Therefore, during the transformation from intestinal metaplasia towards neoplasia, the significant methylation of promoter region in Reprimo and hMLH1 gene might be an early event during gastric cancer onset.

IHC staining showed an elevated expression rate of Reprimo and hMLH1 proteins in intestinal metaplasia, dysplasia and cancer group (in ascending order). Statistical analysis revealed a positive correlation between methylation rate and tissue protein expression rate. As tumor cell metabolites may enter into the plasma during the process of necrosis and apoptosis, the assay of plasma gene methylation level thus may provide a novel non-invasive method for tumor diagnosis with small sample size and simple procedures.

The abnormal methylation of single gene cannot determine the tumor progression. We thus used a combined assay scenario including Reprimo and hMLH1 genes’ methylation status. This method has been shown to own significant value for early diagnosis of gastric cancer. As in either of dysplasia and tumor tissue samples, both genes have been found to have methylation at various levels. The combined assay for methylation owned the positive rate as high as 76% and 84%. Therefore, the combined assay of tissue and plasma levels of Reprimo and hMLH1 gene, can work as one clinical index for the early diagnosis of gastric cancer.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Xiaofeng Yang, Department of Emergency Surgery, Lin-Yi People’s Hospital, 27 Jiefang Road, Lin-Yi 276001, Shangdong Province, China. Tel: +86-539-8110790; Fax: +86-539-8110790; E-mail: xiaofegnyagn@sina.com

References


Gene methylation in gastric cancer


