

Original Article

MicroRNAs and regulated interaction networks reveal differences between adult and pediatric acute myeloid leukemia

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Abstract: Objectives: The purpose of this study was to identify featured microRNAs and their regulated network between adult and pediatric acute myeloid leukemia (AML) and find potential utility as biomarkers for diagnosis and treatment of pediatric AML. Methods: We downloaded the microRNA expression dataset GSE35320 from Gene Expression Omnibus database and selected expression chips from bone marrow of 71 pediatric AML samples and 6 adulthood AML samples. Differentially expressed microRNAs were identified by Wilcox test. The target genes of these microRNAs were predicted using an integrative method and their functional enrichment analysis was performed using DAVID. Finally, STRING database and Cytoscape software was used to construct and analyze the interaction network. Results: A total of 7 differentially expressed microRNAs were identified and the remarkably up-regulated and down-regulated microRNAs were miR-16 and miR-142-5p which included 323 and 22 predicted target genes, respectively. The target genes of 7 microRNAs were most associated with regulation of cell cycle, p53 signaling pathway, Wnt signaling pathway and neurotrophin signaling pathway. The interaction network of miR-16 target genes was constructed among 354 high confidence interaction pairs. The core genes of the network, such as TP53, BCL2, VEGFA, had a role in prognosis of children with AML. Conclusions: The featured microRNAs and their target genes are significant in the occurrence and development of pediatric AML, which is likely to be important for the identification of therapeutic targets and biomarkers for these patients.

Keywords: Pediatric acute myeloid leukemia, microRNA, differential expression, interaction network, miRNA-targeted therapy

Introduction

Acute myeloid leukemia (AML) is a hematologic malignancy that is the second most common type of leukemia in children and accounts for 15-20% of leukemia in children [1]. With advances in chemotherapy and hematopoietic stem cell transplantation, treatment outcome and prognosis of pediatric AML have been greatly improved and the overall survival (OS) rates are now approaching approximately 70% [2, 3]. However, the prognosis for certain high-risk groups of AML and relapsed disease remains poor. There is still considerable need for novel, targeted therapies to further improve the treatment outcome of this disease.

MicroRNAs (miRNAs), a class of about 22 nucleotide-long noncoding RNAs, regulate gene expression at post-transcriptional level and have been predicted to silence over 60% of mammalian genes [4]. They are involved in a variety of critical biological processes and show aberrant expression patterns and functional abnormalities in cancers. Their expression pattern could be used as diagnostic and prognostic biomarkers, as well as oncogenic (or tumor-suppressive) molecules [5]. In adult AML, the roles of microRNAs have been frequently reported. For instance, the miR-29 family that were down-regulated in adult AML samples could function as tumor suppressors, thus their target genes Akt2 and CCND2 that are involved

MicroRNA analyses in ALL

Table 1. Reported microRNAs in pediatric AML

microRNA	Expression pattern	Function	Reference
miR-100	Up	Prognostic biomarker	Bai et al. [25]
miR-125b	Up	Contribute to leukemogenesis and increases drug resistance in pediatric acute promyelocytic leukemia.	Zhang et al. [34]
miR-155, miR-196b	Up	Diagnostic biomarker	Yan et al. [24]
miR-155	Up	Diagnostic and prognostic biomarker	Xu et al. [35]
miR-181a	Up	Regulate G1/S transition and cell proliferation by regulating ATM,	Liu et al. [36]
miR-335	Up	Biomarker for monitoring progression and predicting the clinical outcome	Lin et al. [37]
miR-375	Up	Prognostic biomarker	Wang et al. [38]
miR-99a	Up	Oncogenic microRNA via regulating tumor suppressors CTDSPL and TRIB2	Zhang et al. [39]
miR-9	Down	Tumor suppressor	Emmrich et al. [40]
miR-139-5p	Down	Tumor suppressor in AML by controlling protein translation	Emmrich et al. [41]
miR-370	Down	Diagnostic and prognostic biomarker	Lin et al. [42]
miR-663	Down	Hyper-methylation of the miR-663 promoter may be an early event in the development of pediatric AML	Yan et al. [43]
miR-126, -146a, -181a/b, -100, and miR-125b	-	Distinguish cytogenetic subgroups in pediatric AML	Daschkey et al. [44]
miR-128a, -128b, -223, and let-7b	-	Discriminate ALL from AML cases	Mi et al. [45]
miR-100, miR-125b, miR-335, miR-146a, and miR-99a	-	Highly expressed in pediatric AML when compared with ALL	Zhang et al. [46]

ALL: Acute lymphoblastic leukemia.

Table 2. Differentially expressed microRNAs (P<0.05 and |FC|>1.5)

microRNA	log (Fold change)	p-value	Targets
miR-16	3.50	0.0450	323
miR-221	2.04	0.0112	162
miR-10a	1.10	0.0002	191
let-7b	1.03	0.0026	137
miR-92a	0.93	0.0080	169
miR-26b	-0.86	0.0020	525
miR-142-5p	-4.40	0.0005	22

in the regulation of cell proliferation and cell apoptosis were up-regulated [6]. Low expression level of miR-193b represses AML cell proliferation [7]. Garzon et al. demonstrated that several microRNA signatures were associated with cytogenetic and molecular abnormalities in AML and high expression of miR-191 and miR-199a were significantly correlated with poor prognosis [8]. In pediatric AML, there also have been some related studies about microRNAs, but it still remains limited. As shown in **Table 1**, we have summarized the microRNAs that play important roles in human pediatric AML. However, in both the diagnostic criteria and disease management, there are important differences between adults and children with AML [3]. So understanding the differences in microRNAs profiling between adults and pediatric AML will expand the potential of diagnostics and promote the identification of key microRNAs and genes, providing better insight into the leukemogenesis of pediatric AML. The aim of the present study was to identify important microRNAs and their regulated interaction network between adults and children with AML.

Material and methods

MicroRNA data and data analysis

The microRNA high-throughput dataset GSE-35320 that contains AML samples from cell lines, bone marrow and pediatric patients and adult patients were download from Gene Expression Omnibus database [9]. In order to better compare the microRNAs expression between pediatric and adult patients, we just selected the chips that were all from bone marrow. In total, the processed expression data from 71 pediatric AML patient samples and 6 adulthood AML samples were used. The annotation information of the chips was available based on the

GPL10993 platform. Differences between pediatric and children AML samples were calculated using Wilcox test and microRNAs with fold difference >1.5 and P<0.05 were considered statistically significant.

Prediction the target gens of the differentially expressed microRNAs

To obtain the target genes of the differentially expressed microRNAs, we employed a method [10] that integrates experimentally validated information from miRecords [11], TarBase [12], miR2Disease [13], and miRTarBase [14], and computational prediction results that were predicted at least two tools from HOCTAR [15], ExprTarget [16] and starBase [17].

Functional enrichment analysis

Gene ontology (GO) and KEGG pathway enrichment analysis of the target genes were performed using Database for Annotation, Visualization and Integrated Discovery (DAVID) [18]. The terms with Q-value (Benjamini) <0.01 were selected as highly significantly enriched.

Network construction and analysis

The Search Tool for the Retrieval of the Interacting Genes (STRING) [19], which provided both experimental and predicted interaction information, was used to search and construct the interaction network of the target genes of the differentially expressed microRNAs. The pairs with a combination score >0.7 were selected. Finally, the interaction network was analyzed and the degree of each node was calculated by Cytoscape. The core of the network was defined as the top 10 nodes whose degree is larger than the average degree in the network.

Results

Screening differentially expressed microRNAs

To identify the differentially expressed microRNAs between adults and children AML patients, publicly microarray datasets GSE35320 were obtained from GEO database. At a P<0.05 and |Fold Change|>1.5, a total of 7 microRNAs was considered to be differentially expressed between adults and children AML patient samples, including 5 upregulated and 2 downregulated microRNAs (**Table 2**). miR-16 and miR-

MicroRNA analyses in ALL

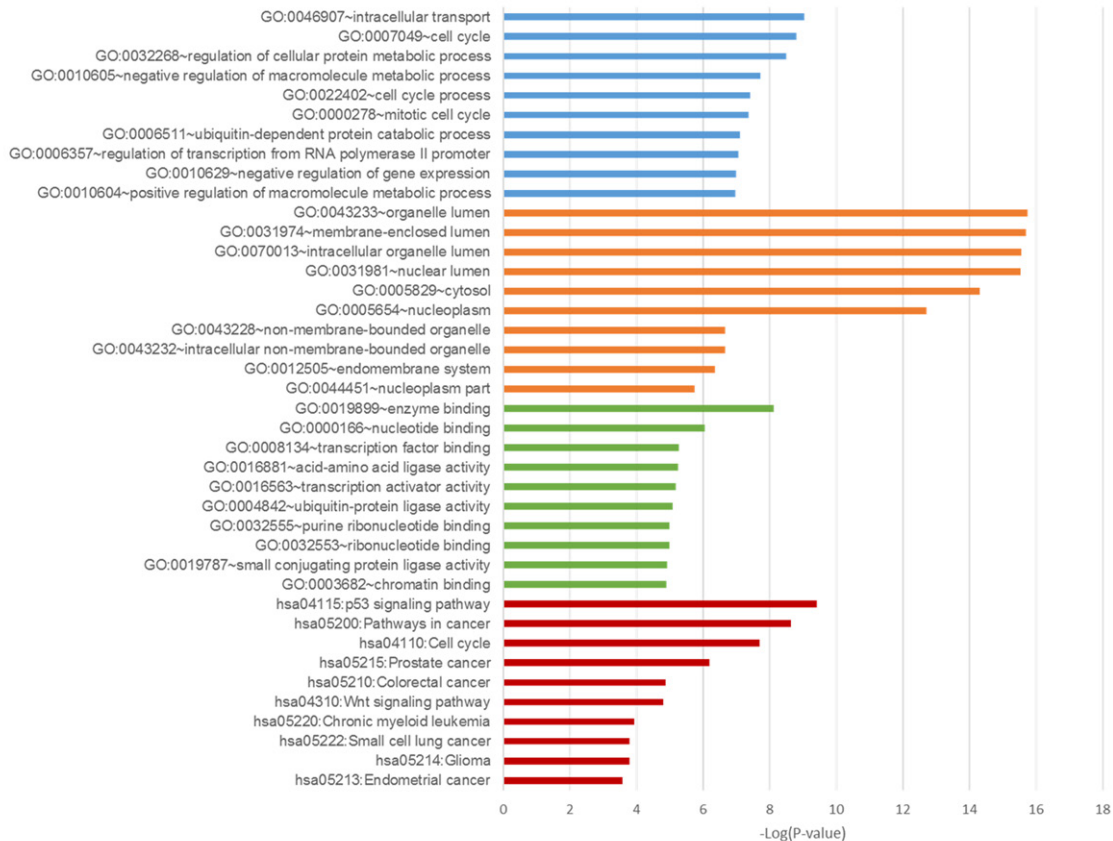


Figure 1. The Top 10 significant enriched DAVID terms in three GO domains and KEGG of target genes. Biological process, cellular component and molecular function terms are marked by blue, orange bars and green bars. KEGG pathways are marked by red bars.

142-5p exhibited the greatest upregulation and downregulation, separately.

Prediction target genes of differentially expressed microRNAs and functional enrichment analysis

The target genes of differentially expressed microRNAs were predicted by an integration method that was elaborated in the methods and materials. The number of target genes of each microRNA was shown in **Table 2**. miR-26b and miR-16 had the top two most predicted target genes. All the predicted target genes of the differentially expressed microRNAs were listed in [Supplemental File 1](#).

To explore the function and annotation of the target genes, the GO that covers three domains: biological process, cellular component and molecular function and pathway enrichment analysis was performed by DAVID (**Figure 1** and

[Supplemental File 2](#)). In biological process domain (blue bar), 87 terms were significantly enriched and many of them related regulation and cell cycle. There were 19 terms, many of which related with lumen and membrane, significantly enriched in cellular component. In molecular function domains, 21 terms were significantly enriched.

Additionally, 11 KEGG pathways were significantly enriched by the target genes of differentially expressed microRNAs. It is notable that all of the significantly enriched pathways were related with cancer and three of them directly with AML including p53 signaling pathway [20, 21], Wnt signaling pathway [22] and neurotrophin signaling pathway [23].

Construction and analysis of interaction network

The target genes of miR-142-5p and miR-16 were mapped to STRING database to screen

MicroRNA analyses in ALL

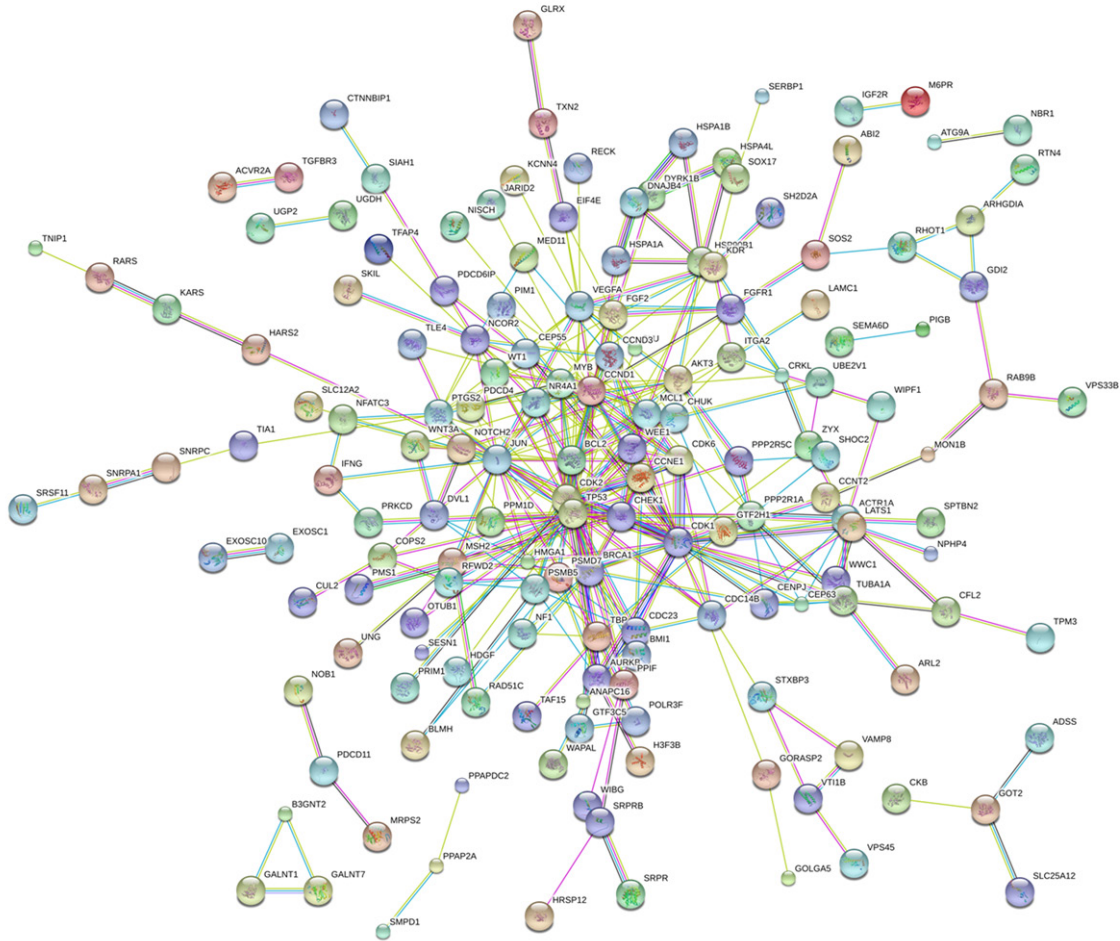


Figure 2. Interaction network of miR-16 target genes constructed by String.

significant interactions and construct their interaction networks. In the condition of combined score >0.7 , there were no interactions among the target genes of miR-142-5p. Among 323 target genes of miR-16, 354 high confidence pairs of interactions were constructed as shown in **Figure 2** and **Supplemental File 3**. The top ten hub genes including TP53 (tumor protein p53), CDK1 (cyclin-dependent kinase 1), CCND1 (cyclin D1), JUN (jun proto-oncogene), CDK2 (cyclin-dependent kinase 2), BCL2 (B-cell CLL/lymphoma 2), VEGFA (vascular endothelial growth factor A), BRCA1 (breast cancer 1), MYB (v-myb myeloblastosis viral oncogene homolog) and FGF2 (fibroblast growth factor 2) constitute the core of the interaction network of miR-16 targets. Most of the core genes were related with cancer, suggesting the important role of miR-16 in pediatric AML.

Discussion

AML is one of the most common types of leukemia in children and the prognosis of pediatric AML is still poor. Therefore, there is an urgent requirement to investigate the mechanism of pediatric AML and to develop an effective preventative strategy specific for children. Recently, it has been demonstrated that microRNAs may represent molecular biomarkers for pediatric patients with AML [24, 25].

In the present study, the gene expression profile GSE35320 from GEO was used to analyze the differentially expressed microRNAs between adults and pediatric AML. As a result, seven microRNAs were obtained, as well as their target genes. Then the functional enrichment analysis was performed for the targets of selected microRNAs and quite a few of the significantly enriched biological processes were

related with cancer, especially three significantly enriched pathways directly play an important role in AML, which suggests that the differentially expressed microRNAs also have a role in pediatric AML.

Finally, the interaction networks of the miR-16 (most increased) and miR-142-5p (most decreased) target genes. Some of the core genes of the interaction network of miR-16 targets can be the biomarkers for pediatric AML. The high level protein expression of p53 can be a potential predictor of early relapse after hematopoietic stem cell transplantation in children with AML [21]. The expression BCL2 is an important prognostic feature to chemotherapy in de novo AML [26] and the relationships and relative levels with other proteins such as tyrosine kinase (FLT3) [27] can predict inferior outcome in pediatric AML. High levels of VEGFA correlated with overall survival [28] and it induced cell survival, proliferation, and protection against apoptosis via VEGFR signaling in pediatric AML [29, 30]. Although no interactions were detected among the target genes of miR-142-5p, the targets themselves were related with AML. For example, overexpression of ATP1B1 was associated with shorter overall survival (OS) and event-free survival of AML patients [31]. Patients with SF3B1 mutation presented with lower hemoglobin levels, increased platelet counts, and have shorter disease-free survival and overall survival than those without the mutation [32, 33]. These results demonstrated the potential roles of miR-16 and miR-142-5p and their targets in pathological mechanism of pediatric AML and can be potentially biomarkers for pediatric AML.

In conclusion, seven microRNAs were identified by comparing adults and children AML bone marrow samples and the targeted genes regulated by microRNAs indicated that they were correlated with p53 signaling pathway, Wnt signaling pathway and neurotrophin signaling pathway. miR-16 and miR-142-5p are likely to be presented as novel potential therapeutic targets and biomarkers for pediatric patients with AML. Further analyses and validation experiment are required to unravel the involvement pediatric AML development.

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Disclosure of conflict of interest

None.

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References

- [1] Seth R and Singh A. Leukemias in Children. *Indian J Pediatr* 2015; 82: 817-24.
- [2] Taga T, Tomizawa D, Takahashi H, Adachi S. Acute myeloid leukemia in children: Current status and future directions. *Pediatr Int* 2016; 58: 71-80.
- [3] Creutzig U, van den Heuvel-Eibrink MM, Gibson B, Dworzak MN, Adachi S, de Bont E, Harbott J, Hasle H, Johnston D, Kinoshita A, Lehrnbecher T, Leverger G, Mejstrikova E, Meshinchi S, Pession A, Raimondi SC, Sung L, Stary J, Zwaan CM, Kaspers GJ, Reinhardt D; AML Committee of the International BFM Study Group. Diagnosis and management of acute myeloid leukemia in children and adolescents: recommendations from an international expert panel. *Blood* 2012; 120: 3187-205.
- [4] Friedman RC, Farh KK, Burge CB, Bartel DP. Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res* 2009; 19: 92-105.
- [5] Lee YS and Dutta A. MicroRNAs in cancer. *Annu Rev Pathol* 2009; 4: 199-227.
- [6] Gong JN, Yu J, Lin HS, Zhang XH, Yin XL, Xiao Z, Wang F, Wang XS, Su R, Shen C, Zhao HL, Ma YN, Zhang JW. The role, mechanism and potentially therapeutic application of microRNA-29 family in acute myeloid leukemia. *Cell Death Differ* 2014; 21: 100-12.
- [7] Gao XN, Lin J, Gao L, Li YH, Wang LL, Yu L. MicroRNA-193b regulates c-Kit proto-oncogene and represses cell proliferation in acute myeloid leukemia. *Leuk Res* 2011; 35: 1226-32.
- [8] Garzon R, Volinia S, Liu CG, Fernandez-Cymering C, Palumbo T, Pichiorri F, Fabbri M, Coombes K, Alder H, Nakamura T, Flomenberg N, Marcucci G, Calin GA, Kornblau SM, Kantarjian H, Bloomfield CD, Andreeff M, Croce CM. MicroRNA signatures associated with cytogenetics and prognosis in acute myeloid leukemia. *Blood* 2008; 111: 3183-9.
- [9] Edgar R, Domrachev M and Lash AE. Gene Expression Omnibus: NCBI gene expression and hybridization array data repository. *Nucleic Acids Res* 2002; 30: 207-10.
- [10] Zhang W, Zang J, Jing X, Sun Z, Yan W, Yang D, Shen B, Guo F. Identification of candidate miRNA biomarkers from miRNA regulatory network

MicroRNA analyses in ALL

- with application to prostate cancer. *J Transl Med* 2014; 12: 66.
- [11] Xiao F, Zuo Z, Cai G, Kang S, Gao X, Li T. miRecords: an integrated resource for microRNA-target interactions. *Nucleic Acids Res* 2009; 37: D105-10.
- [12] Sethupathy P, Corda B and Hatzigeorgiou AG. TarBase: A comprehensive database of experimentally supported animal microRNA targets. *RNA* 2006; 12: 192-7.
- [13] Jiang Q, Wang Y, Hao Y, Juan L, Teng M, Zhang X, Li M, Wang G, Liu Y. miR2Disease: a manually curated database for microRNA deregulation in human disease. *Nucleic Acids Res* 2009; 37: D98-104.
- [14] Hsu SD, Lin FM, Wu WY, Liang C, Huang WC, Chan WL, Tsai WT, Chen GZ, Lee CJ, Chiu CM, Chien CH, Wu MC, Huang CY, Tsou AP, Huang HD. miRTarBase: a database curates experimentally validated microRNA-target interactions. *Nucleic Acids Res* 2011; 39: D163-9.
- [15] Gennarino VA, Sardiello M, Avellino R, Meola N, Maselli V, Anand S, Cuttillo L, Ballabio A, Banfi S. MicroRNA target prediction by expression analysis of host genes. *Genome Res* 2009; 19: 481-90.
- [16] Gamazon ER, Im HK, Duan S, Lussier YA, Cox NJ, Dolan ME, Zhang W. Exptarget: an integrative approach to predicting human microRNA targets. *PLoS One* 2010; 5: e13534.
- [17] Yang JH, Li JH, Shao P, Zhou H, Chen YQ, Qu LH. starBase: a database for exploring microRNA-mRNA interaction maps from Argonaute CLIP-Seq and Degradome-Seq data. *Nucleic Acids Res* 2011; 39: D202-9.
- [18] Huang da W, Sherman BT and Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc* 2009; 4: 44-57.
- [19] Szklarczyk D, Franceschini A, Kuhn M, Simonovic M, Roth A, Minguez P, Doerks T, Stark M, Muller J, Bork P, Jensen LJ, von Mering C. The STRING database in 2011: functional interaction networks of proteins, globally integrated and scored. *Nucleic Acids Res* 2011; 39: D561-8.
- [20] Megonigal MD, Rappaport EF, Nowell PC, Lange BJ, Felix CA. Potential role for wild-type p53 in leukemias with MLL gene translocations. *Oncogene* 1998; 16: 1351-6.
- [21] Mattsson K, Honkaniemi E, Barbany G, Gustafsson B. Increased p53 protein expression as a potential predictor of early relapse after hematopoietic stem cell transplantation in children with acute myelogenous leukemia. *Pediatr Transplant* 2015; 19: 767-75.
- [22] Cheng CK, Li L, Cheng SH, Ng K, Chan NP, Ip RK, Wong RS, Shing MM, Li CK, Ng MH. Secreted-frizzled related protein 1 is a transcriptional repression target of the t(8;21) fusion protein in acute myeloid leukemia. *Blood* 2011; 118: 6638-48.
- [23] Eguchi M, Eguchi-Ishimae M, Tojo A, Morishita K, Suzuki K, Sato Y, Kudoh S, Tanaka K, Setoyama M, Nagamura F, Asano S, Kamada N. Fusion of ETV6 to neurotrophin-3 receptor TRKC in acute myeloid leukemia with t(12;15)(p13;q25). *Blood* 1999; 93: 1355-63.
- [24] Yan W, Xu L, Sun Z, Lin Y, Zhang W, Chen J, Hu S, Shen B. MicroRNA biomarker identification for pediatric acute myeloid leukemia based on a novel bioinformatics model. *Oncotarget* 2015; 6: 26424-36.
- [25] Bai J, Guo A, Hong Z, Kuai W. Upregulation of microRNA-100 predicts poor prognosis in patients with pediatric acute myeloid leukemia. *Onco Targets Ther* 2012; 5: 213-9.
- [26] Maung ZT, MacLean FR, Reid MM, Pearson AD, Proctor SJ, Hamilton PJ, Hall AG. The relationship between bcl-2 expression and response to chemotherapy in acute leukaemia. *Br J Haematol* 1994; 88: 105-9.
- [27] Sharawat SK, Bakhshi R, Vishnubhatla S, Bakhshi S. High receptor tyrosine kinase (FLT3, KIT) transcript versus anti-apoptotic (BCL2) transcript ratio independently predicts inferior outcome in pediatric acute myeloid leukemia. *Blood Cells Mol Dis* 2015; 54: 56-64.
- [28] de Bont ES, Fidler V, Meeuwssen T, Scherpen F, Hählen K, Kamps WA. Vascular endothelial growth factor secretion is an independent prognostic factor for relapse-free survival in pediatric acute myeloid leukemia patients. *Clin Cancer Res* 2002; 8: 2856-61.
- [29] Gerber HP, Malik AK, Solar GP, Sherman D, Liang XH, Meng G, Hong K, Marsters JC, Ferrara N. VEGF regulates haematopoietic stem cell survival by an internal autocrine loop mechanism. *Nature* 2002; 417: 954-8.
- [30] Santos SC and Dias S. Internal and external autocrine VEGF/KDR loops regulate survival of subsets of acute leukemia through distinct signaling pathways. *Blood* 2004; 103: 3883-9.
- [31] Shi JL, Fu L, Ang Q, Wang GJ, Zhu J, Wang WD. Overexpression of ATP1B1 predicts an adverse prognosis in cytogenetically normal acute myeloid leukemia. *Oncotarget* 2016; 7: 2585-95.
- [32] Hou HA, Liu CY, Kuo YY, Chou WC, Tsai CH, Lin CC, Lin LI, Tseng MH, Chiang YC, Liu MC, Liu CW, Tang JL, Yao M, Li CC, Huang SY, Ko BS, Hsu SC, Chen CY, Lin CT, Wu SJ, Tsay W, Tien HF. Splicing factor mutations predict poor prognosis in patients with de novo acute myeloid leukemia. *Oncotarget* 2016; 7: 9084-101.
- [33] Damm F, Kosmider O, Gelsi-Boyer V, Renneville A, Carbuccia N, Hidalgo-Curtis C, Della Valle V, Couronné L, Scourzic L, Chesnais V, Guerci-Bresler A, Slama B, Beyne-Rauzy O, Schmidt-

MicroRNA analyses in ALL

- Tanguy A, Stamatoullas-Bastard A, Dreyfus F, Prébet T, de Botton S, Vey N, Morgan MA, Cross NC, Preudhomme C, Birnbaum D, Bernard OA, Fontenay M; Groupe Francophone des Myélo-dysplasies. Mutations affecting mRNA splicing define distinct clinical phenotypes and correlate with patient outcome in myelodysplastic syndromes. *Blood* 2012; 119: 3211-8.
- [34] Zhang H, Luo XQ, Feng DD, Zhang XJ, Wu J, Zheng YS, Chen X, Xu L, Chen YQ. Upregulation of microRNA-125b contributes to leukemogenesis and increases drug resistance in pediatric acute promyelocytic leukemia. *Mol Cancer* 2011; 10: 108.
- [35] Xu LH, Guo Y, Cen JN, Yan WY, He HL, Niu YN, Lin YX, Chen CS, Hu SY. Overexpressed miR-155 is associated with initial presentation and poor outcome in Chinese pediatric acute myeloid leukemia. *Eur Rev Med Pharmacol Sci* 2015; 19: 4841-50.
- [36] Liu X, Liao W, Peng H, Luo X, Luo Z, Jiang H, Xu L. miR-181a promotes G1/S transition and cell proliferation in pediatric acute myeloid leukemia by targeting ATM. *J Cancer Res Clin Oncol* 2016; 142: 77-87.
- [37] Lin X, Wang Z, Zhang R, Feng W. High serum microRNA-335 level predicts aggressive tumor progression and unfavorable prognosis in pediatric acute myeloid leukemia. *Clin Transl Oncol* 2015; 17: 358-64.
- [38] Wang Z, Hong Z, Gao F, Feng W. Upregulation of microRNA-375 is associated with poor prognosis in pediatric acute myeloid leukemia. *Mol Cell Biochem* 2013; 383: 59-65.
- [39] Zhang L, Li X, Ke Z, Huang L, Liang Y, Wu J, Zhang X, Chen Y, Zhang H, Luo X. MiR-99a may serve as a potential oncogene in pediatric myeloid leukemia. *Cancer Cell Int* 2013; 13: 110.
- [40] Emmrich S, Katsman-Kuipers JE, Henke K, Khatib ME, Jammal R, Engeland F, Dasci F, Zwaan CM, den Boer ML, Verboon L, Stary J, Baruchel A, de Haas V, Danen-van Oorschot AA, Fornerod M, Pieters R, Reinhardt D, Klusmann JH, van den Heuvel-Eibrink MM. miR-9 is a tumor suppressor in pediatric AML with t(8;21). *Leukemia* 2014; 28: 1022-32.
- [41] Emmrich S, Katsman-Kuipers JE, Henke K, Khatib ME, Jammal R, Engeland F, Dasci F, Zwaan CM, den Boer ML, Verboon L, Stary J, Baruchel A, de Haas V, Danen-van Oorschot AA, Fornerod M, Pieters R, Reinhardt D, Klusmann JH, van den Heuvel-Eibrink MM. miR-139-5p controls translation in myeloid leukemia through EIF4G2. *Oncogene* 2016; 35: 1822-31.
- [42] Lin X, Wang Z, Wang Y, Feng W. Serum MicroRNA-370 as a potential diagnostic and prognostic biomarker for pediatric acute myeloid leukemia. *Int J Clin Exp Pathol* 2015; 8: 14658-66.
- [43] Yan-Fang T, Jian N, Jun L, Na W, Pei-Fang X, Wen-Li Z, Dong W, Li P, Jian W, Xing F, Jian P. The promoter of miR-663 is hypermethylated in Chinese pediatric acute myeloid leukemia (AML). *BMC Med Genet* 2013; 14: 74.
- [44] Daschkey S, Röttgers S, Giri A, Bradtke J, Teigler-Schlegel A, Meister G, Borkhardt A, Landgraf P. MicroRNAs distinguish cytogenetic subgroups in pediatric AML and contribute to complex regulatory networks in AML-relevant pathways. *PLoS One* 2013; 8: e56334.
- [45] Mi S, Lu J, Sun M, Li Z, Zhang H, Neilly MB, Wang Y, Qian Z, Jin J, Zhang Y, Bohlander SK, Le Beau MM, Larson RA, Golub TR, Rowley JD, Chen J. MicroRNA expression signatures accurately discriminate acute lymphoblastic leukemia from acute myeloid leukemia. *Proc Natl Acad Sci U S A* 2007; 104: 19971-6.
- [46] Zhang H, Luo XQ, Zhang P, Huang LB, Zheng YS, Wu J, Zhou H, Qu LH, Xu L, Chen YQ. MicroRNA patterns associated with clinical prognostic parameters and CNS relapse prediction in pediatric acute leukemia. *PLoS One* 2009; 4: e7826.