Original Article

Immunohistochemical expression of nuclear and cytoplasmic survivin in gastrointestinal carcinoma

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Abstract: Survivin is a protein that is highly expressed in many embryonic tissues, as well as most human tumors. Prior studies have reported both positive and negative correlations between survivin expression and cancer prognosis, but these associations remain controversial. In the present study, we assessed the expression of nuclear and cytoplasmic survivin in gastrointestinal carcinomas. Using these data, we determined the correlation between nuclear and cytoplasmic survivin and, further, investigated correlations between survivin expression and clinicopathological parameters. Seventy-two advanced gastric adenocarcinomas and 78 colorectal adenocarcinomas were analyzed for survivin expression by immunohistochemistry. Expression of both nuclear and cytoplasmic survivin was significantly higher in colorectal carcinomas than in gastric carcinomas (P<0.01). There was a positive correlation between nuclear and cytoplasmic expression of survivin (r=0.42, P<0.001). In gastric carcinomas, the level of survivin protein expression was associated with tumor differentiation, patient age, and lymphatic invasion (P<0.05, 0.01, and 0.01, respectively). In colorectal carcinomas, the level of nuclear survivin expression was significantly higher in females than in males (P<0.05). There were no significant associations between survivin expression and most of the clinicopathological parameters. Nevertheless, there was a trend towards an inverse correlation between nuclear survivin expression and tumor aggressiveness in gastric carcinoma; there was a similar trend for cytoplasmic survivin expression. In summary, our results suggest that levels of nuclear and cytoplasmic survivin expression differ between gastric carcinoma and colorectal carcinoma.

Keywords: Survivin, immunohistochemistry, gastrointestinal carcinoma, apoptosis

Introduction

Survivin, a member of the inhibitor of apoptosis protein (IAP) family, has a dual cellular function as an inhibitor of apoptosis and as a regulator of mitosis. Its anti-apoptotic function is related to its inhibition of caspase activity by directly or indirectly interfering with the function of the caspase-3, caspase-7, and caspase-9 [1-4]. In addition to its anti-apototic function, survivin functions as a chromosome passenger protein, regulating the G2 and M phases of the cell cycle. The chromosomal passenger complex consists of Aurora B kinase, INCENP, survivin, and Borealin [5-8].

Survivin is highly expressed in many embryonic tissues, as well as most human tumors of the lung, colon, breast, stomach, liver, ovary, and prostate. In contrast, it is either undetectable or expressed at a very low level in differentiated adult tissues [9-19]. Therefore, it has been suggested that survivin could be a marker for tumor progression and prognosis [20-32]. Furthermore, given its functional properties, survivin has been proposed as a molecular target for anti-cancer therapies [12, 33-37]. However, the molecular mechanisms regulating survivin remain poorly understood.

Survivin localizes to the nucleus and cytoplasm of cancer cells. Several studies have proposed that the subcellular distribution of survivin is regulated by active import into the nucleus and CRM1-mediated export to the cytoplasm [7, 37, 38]. It has also been suggested that survivin could be a nuclear shuttling protein. Recent studies have suggested that the nuclear pool of
survivin is involved in promoting cell proliferation, whereas the cytoplasmic pool of survivin controls cell survival [7, 28, 37].

Previous studies have reported both positive and negative correlations between survivin expression and cancer prognosis, but these results remain controversial (Tables 1 and 2). Several reports have suggested that nuclear expression of survivin is a prognostic marker. Some reports have demonstrated an association between nuclear expression of survivin and unfavorable outcomes in patients with gastric and colorectal carcinomas [23]. Conversely, other reports have demonstrated that nuclear expression of survivin is positively correlated with favorable prognoses in gastric and colorectal carcinomas [22, 28]. Finally, other reports...
have demonstrated that cytoplasmic survivin expression is correlated with better, worse, or unchanged prognoses in patients with gastric and colorectal carcinomas [20, 27-29, 33].

To date, no study has investigated the relationship between expression of nuclear survivin and cytoplasmic survivin. Therefore, the present study evaluated expression of nuclear and cytoplasmic survivin in gastrointestinal carcinomas in order to determine their correlation and, furthermore, to identify the relationship between survivin expression and clinicopathological parameters. Our results indicate that gastric carcinomas and colorectal carcinomas differ in levels of expression of nuclear and cytoplasmic survivin.

### Materials and methods

#### Tissue samples

Seventy-two advanced gastric adenocarcinomas (36 well-to-moderately differentiated and 36 poorly differentiated) and 78 colorectal adenocarcinomas (68 well-to-moderately differentiated and 10 poorly differentiated) were analyzed for survivin expression by immunohistochemistry. Surgically resected tumor tissues were collected from the archives of the Department of Diagnostic Pathology of the Osaka Red Cross Hospital and the Kobe Central Hospital of Social Insurance. The study was approved by the local ethics committee. The tumors were classified according to the TNM classification of malignant tumors (TNM 2009) [39]. All specimens were preserved in 10% formalin and embedded in paraffin. Three-micrometer-thick sections were cut consecutively and mounted on amino-propyltriethoxysilane-coated slides.

#### Immunohistochemical staining

The sections were deparaffinized with xylene and rehydrated with a graduated series of ethanol solutions. Endogenous peroxidase was blocked by incubating the sections in 0.03% hydrogen peroxide in methanol for 30 min. Antigen retrieval was performed by immersing slides in 0.1 M citrate buffer (pH 7.0) and heating for 10 minutes in a pressure cooker (T-FAL; Rumily, France). After heating, the sections were cooled at room temperature in the soaking solution for 30 min. The sections were washed under running tap water followed by 0.01 M phosphate buffered saline (PBS; pH 7.2), and then incubated with a rabbit polyclonal antibody against survivin (1:1,000 dilution, R&D Systems Minneapolis, MN, USA) overnight at room temperature. The sections were then rinsed with PBS. To detect survivin, the sections were incubated with the Histofine Simple Stain MAX-PO (Nichirei, Tokyo, Japan) for 1 h at room temperature. The reaction products were developed with dianisobenzidine
and counterstained with Mayer's hematoxylin. As a negative control, a section was treated as described above, but with the primary antibody replaced by buffer.
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Immunostaining evaluation

Sections were considered positive for survivin in the presence of nuclear or cytoplasmic staining. The mean percentage of positive tumor cells was determined using the average of at least five areas at 400× magnification. Samples with ≥ 30% of tumor cells positive for nuclear staining and ≥ 15% of tumor cells positive for cytoplasmic staining were considered to have positive survivin protein expression. These cut-off values of 30% and 15% for nuclear and cytoplasmic staining, respectively, were based on the median observed values. The survivin-stained sections were examined by two independent researchers blinded to other pathological information.

Statistical analysis

The chi-square test and Fisher’s exact test were used to determine statistical differences in marker expression between gastric adenocarcinomas and colorectal adenocarcinomas, well-to-moderately differentiated and poorly-differentiated adenocarcinomas, patient age and gender, and lymphatic invasion and vascular invasion. Correlations between marker expression and tumor location, depth of invasion, lymph node metastasis, or pathological stage were analyzed using the Kruskal-Wallis test. Spearman rank correlation was calculated to assess the correlation between nuclear survivin expression and cytoplasmic survivin expression. P-values of < 0.05 were considered statistically significant.

Results

The clinicopathological parameters of 72 gastric carcinomas and 78 colorectal carcinomas are reported in Table 3.

Expression of survivin in gastric carcinoma

Survivin expression was observed in the nucleus and/or cytoplasm (Figure 1). Survivin-positive nuclear staining was observed in 49% (35/72) of gastric carcinomas, and cytoplasmic survivin expression was detected in 35% (25/72) of gastric carcinomas. Nuclear survivin expression in well-to-moderately differentiated samples (61%) was significantly higher than in poorly differentiated samples (36%; P < 0.05) (Table 4). Cytoplasmic survivin was detected in 31% (11/36) of well-to-moderately differentiat-
ed samples. There was a positive correlation between nuclear and cytoplasmic expression of survivin ($r = 0.42, P < 0.001$).

**Expression of survivin in colorectal carcinoma**

Survivin-positive nuclear staining was observed in 72% (56/78) of colorectal carcinomas, and cytoplasmic survivin expression was detected in 56% (44/78) of colorectal carcinomas. Expression of nuclear and cytoplasmic survivin was significantly higher in colorectal carcinomas than in gastric carcinomas ($P < 0.01$). Nuclear survivin expression was significantly higher than cytoplasmic survivin expression ($P < 0.05$). In contrast to gastric carcinomas, there was no relationship between nuclear survivin expression and cytoplasmic survivin expression in colorectal carcinomas.

**Correlation between survivin expression and clinicopathological parameters**

A clinicopathological analysis of the survivin-positive samples is shown in Figure 2. In gastric carcinomas, the level of survivin protein expression was associated with patient age, and lymphatic invasion ($P < 0.01$, and 0.01, respectively). None of the other parameters (patient gender, tumor location, depth of invasion, lymph-node metastasis, vascular invasion, or pathological stage) was associated with positive survivin expression.

In colorectal carcinomas, the level of nuclear survivin expression was significantly higher in females than in males ($P < 0.05$). None of the other parameters (patient age, tumor location, depth of invasion, lymph-node metastasis, lymphatic invasion, vascular invasion, or pathological stage) was associated with positive survivin expression. Although there were no significant differences between most of the clinicopathological parameters and survivin expression, there was a trend toward an association between decreased nuclear survivin expression and tumor aggressiveness in gastric carcinoma, with cytoplasmic survivin expression exhibiting a similar trend. In contrast, in colorectal carcinomas, cytoplasmic survivin expression increased - equaling or surpassing nuclear survivin expression - with increasing tumor aggressiveness. These data indicate that gastric carcinomas and colorectal carcinomas differ in their patterns of nuclear and cytoplasmic survivin expression.

**Discussion**

In this study, we used immunohistochemistry to investigate subcellular localization of survivin protein in gastric and colorectal carcinomas. Our data indicate that expression of both nuclear and cytoplasmic survivin was significantly higher in colorectal carcinomas (nuclear survivin, 72%; cytoplasmic survivin, 56%) than in gastric carcinomas (nuclear survivin, 49%; cytoplasmic survivin, 35%) ($P < 0.01$). Kawasaki et al. reported a higher incidence of cytoplasmic survivin expression in colorectal carcinomas than in gastric carcinomas (53.2% versus 34.5%) [26]. To our knowledge, ours is the first study comparing both nuclear and cytoplasmic survivin expression between gastric carcinomas and colorectal carcinomas. Furthermore, our results indicate that nuclear survivin expression is significantly higher than cytoplasmic survivin expression ($P < 0.05$) in colorectal carcinomas. This finding is consistent with the report from Qi et al., which suggested that survivin is more highly localized in the nucleus (78%; 109 of 142) than the cytoplasm (20%; 29 of 142) of colorectal carcinomas [28]. Together, these results suggest that the survivin protein is mainly localized in the nucleus rather than in the cytoplasm of colorectal carcinomas.

We next focused on the correlation between survivin expression and tumor differentiation. Among gastric carcinomas, nuclear survivin expression was significantly higher in well-to-moderately differentiated samples (61%) than in poorly differentiated samples (36%) ($P < 0.05$). However, there was no significant difference in cytoplasmic survivin expression between the well-to-moderately differentiated samples (31%) and the poorly differentiated samples (39%). Wakana et al. reported that cytoplasmic survivin expression was significantly higher in diffuse-type cases than in intestinal-type cases of gastric carcinomas [25]. However, Lu et al. used immunohistochemistry to demonstrate that cytoplasmic survivin is more highly localized in the intestinal-type gastric carcinomas than in the diffuse-type gastric carcinomas ($P < 0.05$) [32]. Miyachi et al. reported that survivin mRNA expression was independent of the histological type of gastric cancer [30]. Wakana et al. demonstrated that survivin mRNA expression was significantly higher in diffuse-type gastric carcinomas than in intestinal-type gastric carcinoma [25]. Our
results demonstrate that nuclear survivin protein expression is higher in well-to-moderately differentiated samples than in poorly differentiated samples, and cytoplasmic survivin expression differed from nuclear survivin expression in gastric carcinomas. The discrepancies between the studies are likely due to analyzing survivin mRNA versus survivin protein. Wakana et al. suggested that these discrepancies might be due to differences in tumor cell volume in the tissue specimens or differences in the detection method [25].

We next investigated the relationship between nuclear survivin and cytoplasmic survivin expression. A positive correlation was observed between nuclear and cytoplasmic survivin expression ($r = 0.42, P < 0.001$) in gastric carcinomas; in contrast, no such relationship was observed in colorectal carcinomas. These results indicate that regulation of survivin expression is likely different between gastric carcinomas and colorectal carcinomas.

Finally, we investigated whether there was a correlation between nuclear or cytoplasmic survivin expression and clinicopathological parameters. In gastric carcinomas, survivin expression was correlated with tumor differentiation, patient age, and lymphatic invasion ($P < 0.05, 0.01$, and $0.01$, respectively). Similarly, Lu et al. reported that cytoplasmic survivin was prominent in gastric carcinomas without lymphatic invasion [32]. In addition, we showed that nuclear survivin expression in colorectal carcinomas was significantly higher in females than in males ($P < 0.05$). None of the other parameters was associated with the expression of survivin. Nuclear and cytoplasmic survivin expression tended to decrease with increasing tumor aggressiveness in gastric carcinomas, but this effect was not statistically significant. Similarly, there was an inverse relationship between nuclear survivin expression and cytoplasmic survivin expression in colorectal carcinomas, which did not reach statistical significance.

There are several previous reports on the relationship between prognosis and expression of survivin mRNA or cytoplasmic survivin. Vallböhmer et al. reported that a high level of cytoplasmic survivin expression was associated with significantly higher rates of survival in patients with gastric carcinoma [33]. Similarly, Okada et al. reported that survivin nuclear staining was associated with a favorable prognosis [22]. Ponnelle et al. reported that high levels of survivin expression were associated with increased survival in patients with colorectal carcinomas [27]. On the other hand, Lee et al. reported that a higher cytoplasmic survivin immunostaining score was associated with higher mortality in patients with colorectal carcinoma [29]. Fang et al. reported that elevated expression of survivin was associated with lower rates of survival [21]. Qi et al. reported that higher cytoplasmic survivin expression was associated with a poor prognosis, while higher nuclear survivin expression was associated with a better prognosis [28]. Taken together, these reports indicate that nuclear and cytoplasmic survivin expression is associated with a better prognosis in gastric carcinomas. Conversely, upregulation of cytoplasmic survivin is associated with poor prognosis in colorectal carcinomas.

Survivin localizes to both the nucleus and cytoplasm. Nuclear survivin is thought to promote cell proliferation, and cytoplasmic survivin is thought to have cytoprotective activity [7, 28, 37]. Tu et al. reported that suppression of survivin expression or function in gastric carcinoma led to abnormal morphology, with decreased cell growth and increased rates of spontaneous apoptosis and mitotic catastrophe [24].

In the present study, we found that survivin expression was higher in both the nucleus and cytoplasm of colorectal carcinomas than in gastric carcinomas. Further, we identified a positive correlation between nuclear survivin and cytoplasmic survivin expression in gastric carcinomas but not in colorectal carcinomas. Our previous report suggested that different cell-death pathways are activated in gastric and colorectal carcinomas [40]. The extrinsic and intrinsic apoptotic pathways could be mutually regulated in gastric adenocarcinomas. In contrast, in colorectal carcinomas, autophagy might function as a cellular guardian to avoid caspase 9-dependent apoptosis. The present study suggests that survivin's role in inhibiting apoptosis may be more prominent in colorectal carcinomas than in gastric carcinomas.

Based on our results, we conclude that gastric carcinomas and colorectal carcinomas differ in their levels of nuclear and cytoplasmic survivin.
expression. Although the role of survivin in tumor progression remains unknown, our results suggest that an inhibitor of survivin might be appropriate for individualized treatment of patients with gastric and colorectal cancer.

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Disclosure of conflict of interest

None.

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